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(54) Title: LEAD BINDING POLYPEPTIDES AND NUCLEOTIDES CODING THEREFOR

(57) Abstract

Metal binding polypeptides which include an amino acid sequence coding for a variable region of a monoclonal antibody which immunoreacts with a lead cation and nucleotides which include a nucleic acid sequence coding for the variable region are provided. The invention is also directed to fusion proteins which include the monoclonal antibody heavy chain variable region and a phage coat protein or portion thereof. The invention also provides bacteriophages which include the fusion protein in their coat. In addition, methods for detecting, removing, adding, or neutralizing lead cations in biological or inanimate systems through the use of the lead binding polypeptides are provided. A method of forming a hybridoma which produces monoclonal antibodies capable of immunoreacting with a metallic cation is also provided.

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LFAD BINDING POLYPEPTIDES AND NUCLEOTIDES CODING THEREFORE

Background of the Invention

Small chemical moieties, such as heavy metal ions, can and often do affect the environment and biological These effects become astounding when it is realized that minute quantities of these small moieties are involved. Moreover, the presence or absence of low 10 concentrations of small moieties in the environment can have long term consequences. Minute quantities of metallic cations, such as lead cations, can regulate, influence, change or toxify the environment or biological systems.

The detection, removal, addition or neutralization of such minute quantities constitutes a focal point for continued research in many fields. For example, many efforts have been made to detect and remove minute, toxic amounts of heavy metal ions such as lead or 20 mercury from the environment. The efforts often have not been successful or economical for widespread application. On the other hand, minute concentrations of other heavy metals are important for the proper function of biological organisms. Zinc, for example, 25 plays a major role in wound healing.

Heavy metal can exhibit dual roles. For example, lead is used in glass making and in chemical manufacturing operations. Yet when ingested by mammals, such as from drinking water, lead may be highly toxic in very small amounts. Hence, detection and quantification of minute concentrations of a heavy metal, such as lead, in drinking water and other media would serve exploratory, safety and regulatory goals.

It would, therefore, be highly desirable to identify and control minute quantities of heavy metals, e.g., lead cations, in aqueous biological or inanimate In most contexts, however, the detection, removal, addition or neutralization of heavy metals, is a difficult and expensive and often unfeasible if not

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impossible task. Other metallic contaminants often mimic the heavy metal of interest and result in measurement interference. Moreover, the detection methods employed today are usually not sufficiently sensitive at the minute quantities under consideration. Consequently, it is desirable to develop reliable and economic methods for accurately identifying and controlling minute quantities of a particular heavy metal in the presence of other heavy metals.

Antibodies would seem to be uniquely suited for this task. Their high degree of specificity for a known antigen would avoid the potential interference caused by contaminants. The sensitivity of antibodies in assays to detect analytes in the picomolar or lower range would permit accurate and efficient targeting and detection at minute levels.

Monoclonal antibodies, of course, come to mind as especially suited agents for practice of this technique. Since Kohler and Milstein published their article on the use of somatic cell hybridization to produce monoclonal antibodies (Nature 256:495 (1974)), immunologists have developed many monoclonal antibodies which strongly and specifically immunoreact with antigens.

Notwithstanding this suggestion, the conventional understanding about immunology teaches that antibodies against small moieties, such as heavy metals, cannot be developed. The mammal immunization step, which is key for the production of monoclonal antibodies, typically requires a molecule that is large enough to cause antigenic reaction. Medium sized molecules (haptens), which are not of themselves immunogenic, can induce immune reaction by binding to an immunogenic carrier. Nevertheless, immunologists view small moieties such as metallic cations, as not large or structurally complex enough to elicit an antibody response. The molecular size and lack of complexity of an inorganic cation is

thought to render it insufficient for eliciting an antibody response.

Several immunologists have reported production of monoclonal antibodies to metallic ion chelates. For 5 example, in U.S. Patent No. 4,722,892, monoclonal antibodies are disclosed which immunoreact with a complex of a chelating agent, such as ethylene diamine tetracetate (EDTA), and a heavy metal such as indium. In EPO Patent Application 0235457, monoclonal antibodies 10 that immunoreact with cyanide complexes of gold and/or silver are disclosed. In these instances, however, the monoclonal antibodies bind with the metal chelate complex rather than the bare metallic ion itself. Disadvantages of antigen measurement methods based on 15 such antibodies include: the complicated reagents involved in detection, lack of simple tests that discriminate among antigens, cross-reactivity with chelates of other antigens and cross-reactivity with the chelating agent itself.

Other instances of monoclonal antibody combinations with metals involve metal tags. The metals or metal chelates are bound to the antibody at a site remote from the antigen binding site or sites. The metal tag is not the antigen. Instead, the metal tag is used to indicate the presence of the monoclonal antibody when it reacts with its specific antigen. See for example, V.P. Torchilian et al., Hybridoma, 6, 229 (1987); and C.F. Meares, Nuclear Medical Biology, 13, 311-318 (1986).

Accordingly, there exists a need to develop

30 polypeptides that immunoreact with heavy metals per se
and with lead ions in particular. This would permit the
development of methods for detecting or neutralizing
heavy metals within, adding heavy metals to, or removing
heavy metals from biological or inanimate systems

35 through the use of the monoclonal antibodies. There
also exists a need for the development of nucleic acid

sequences coding for polypeptides which selectively bind

with lead cations and the development of methods of expressing and translating these nucleic acid sequences to produce metal binding polypeptides.

5 Summary of the Invention

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The present invention provides a metal binding . polypeptide which selectively binds a heavy metal, such as a lead cation. The metal binding polypeptide includes an amino acid sequence for a variable region 10 from a monoclonal antibody, wherein the monoclonal antibody immunoreacts with a lead cation. For example, the metal binding polypeptide may include an amino acid sequence for a heavy chain Fd fragment (consisting of the heavy-chain variable region and heavy-chain constant region 1 domains) from the monoclonal antibody. metal binding polypeptide may further include a heavy chain Fc fragment fused to the heavy chain Fd fragment or a phage coat protein or portion thereof fused to the heavy chain Fd fragment. Alternatively, the metal 20 binding polypeptide may include an amino acid sequence for a light chain from the monoclonal antibody.

Another embodiment of the invention provides a recombinantly produced Fab fragment that immunoreacts with a lead cation. The recombinantly produced Fab 25 fragment includes an amino acid sequence for a variable region from a monoclonal antibody which immunoreacts with the lead cation. Preferably, the Fab fragment includes a heavy chain Fd fragment or a light chain from the monoclonal antibody.

The present invention also provides a purified antibody which includes a Fab fragment. The Fab fragment immunoreacts with a lead cation and includes an amino acid sequence selected from a group of sequences for a variable region of certain specified monoclonal antibodies. The Fab fragment heavy chain preferably includes an amino acid sequence selected from a group of the sequences for the heavy chain variable region of the

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specified monoclonal antibodies. In another preferred embodiment, the Fab fragment light chain includes an amino acid sequence selected from a group of the sequences for the light chain variable region of the specified monoclonal antibodies. The monoclonal antibody may be a recombinantly produced monoclonal antibody.

The invention is also directed to a heavy chain of the monoclonal antibody. The heavy chain preferably includes a sequence selected from a group of the sequences for the heavy chain variable region of certain specified monoclonal antibodies. The invention is also directed to a light chain of the monoclonal antibody. As with the heavy chain, the light chain preferably includes a sequence selected from a group of the sequences for the light chain variable region of certain specified monoclonal antibodies.

Yet another embodiment of the invention is directed to an isolated nucleic acid sequence coding for a variable region of a monoclonal antibody, e.g., the heavy chain variable region or the light chain variable region of the monoclonal antibody. The monoclonal antibody immunoreacts with a lead cation.

Alternatively, the isolated nucleic acid sequence may code for the heavy chain Fd fragment, the entire heavy chain or the entire light chain of the monoclonal antibody.

The present invention is also directed to an expression cassette. The expression cassette includes a nucleic acid sequence coding for a variable region of the monoclonal antibody which immunoreacts with a lead cation. The nucleic acid sequence coding for the variable region is operably linked to a promoter functional in a vector. The expression cassette may include the promoter operably linked to a nucleic acid sequence coding for a heavy chain Fd fragment of the monoclonal antibody. Alternatively, the expression

cassette may include the promoter operably linked to a nucleic acid sequence coding for a light chain of the monoclonal antibody. The expression cassette may also include a leader sequence located between the promoter and the nucleic acid sequence coding for the monoclonal antibody chain. The leader sequence may function to direct the heavy or light chain to a membrane in a host cell or to cause the antibody chain to be secreted by the host cell.

In another embodiment, the present invention provides a fusion protein which includes a phage coat protein or portion thereof fused to an amino acid sequence for a heavy chain variable region from the monoclonal antibody. The fusion protein preferably includes the heavy chain Fd fragment of the monoclonal antibody. The fusion protein may be present as part of the coat of a phage and, preferably, the coat of a filamentous phage.

Another embodiment of the present invention is 20 directed to an expression cassette coding for a fusion protein. This expression cassette includes a first nucleic acid sequence coding for a heavy chain Fd fragment of a monoclonal antibody which reacts with a lead cation. The first nucleic acid sequence is linked 25 for co-expression to a second nucleic acid sequence, such as a nucleic acid sequence coding for a phage coat protein or a portion thereof, to form a nucleic acid sequence encoding the fusion protein. The fusion protein includes the heavy chain Fd fragment fused to 30 the phage coat protein or portion thereof. expression cassette coding for the fusion protein also includes a promoter that is functional in a vector. promoter is operably linked to the first and second DNA sequences and provides for expression of the fusion protein. The expression cassette may also include a leader sequence which directs expression of the fusion protein to a membrane of a host cell. The leader

sequence is located between the promoter and the nucleic acid sequence coding for the fusion protein. addition, the expression cassette may include a third nucleic acid sequence coding for a peptide linker. 5 third nucleic acid sequence is typically located between the first and second nucleic acid sequences. expression cassette may optionally include a fourth nucleic acid sequence coding for a light chain of a monoclonal antibody. Preferably, the light chain is a 10 light chain of a monoclonal antibody that immunoreacts with a lead cation. The present invention also provides a phagemid vector which includes one of the expression cassettes described above.

The invention is also directed to methods for 15 detecting, removing, adding, or neutralizing the heavy metals in biological and inanimate systems through the use of the metal binding polypeptides, heavy and light chains, fusion proteins, recombinantly produced Fab fragments and monoclonal antibodies described above.

The advantages of the invention include among others: the lack of complication by additional reagents, a high discrimination against similar metallic cations, lack of cross-reactivity with similar metallic cations, and lack of cross-reactivity with test 25 reagents.

The metal binding polypeptide of the invention binds with a heavy metal per se, and preferably with a lead cation per se. The metal binding polypeptide binds with a heavy metal cation which is at least partially 30 exposed, i.e., a heavy metal cation which is not complexed or enveloped. For example, the heavy metal cation may be associated with a spacer arm which includes a sulfur atom as a Lewis base group. The state of the heavy metal during the immunoreaction is preferably one of non-coordination with any other substance; in other words, bare or exposed.

Preferably, the metal binding polypeptide exhibits a substantially high degree of specific immunoreactivity toward the heavy metal. More preferably, the metal binding polypeptide includes a portion of a recombinantly produced Fab fragment (e.g., the light chain or the heavy chain Fd fragment of the Fab fragment) and has an association constant for a heavy metal such as a lead cation that is about 10,000 fold greater than the association constant for the immunogen compound without the heavy metal. Also preferably, the 10 metal binding polypeptide is immunospecific for a particular member of a group of very similar heavy metals. The monoclonal antibody will typically exhibit a relative association constant for such a particular 15 heavy metal that is about 10,000 fold greater than that for the other heavy metals of such a group.

The hybridoma of the invention, which produces the monoclonal antibody, is formed from immune cells that are specific for the heavy metal. The formation may be 20 accomplished by fusion of an immortal mammal cell line and mammalian immune cells from a second mammal previously immunized with an immunogen compound which contains the heavy metal. A lead cation monoclonal antibody produced by this first type of hybridoma is 25 hereinafter referred to as a "Group 1 lead cation monoclonal antibody." Alternatively, the hybridoma may be formed by fusion of an immortal mammal cell line and mammalian immune cells from a second mammal previously immunized with a second monoclonal antibody capable of 30 immunoreacting with the heavy metal. A lead cation monoclonal antibody produced by this latter type of hybridoma is hereinafter referred to as a "Group 3 lead cation monoclonal antibody." Selection of the appropriate hybridoma may be carried out by cross-35 screening the secreted monoclonal antibody against the heavy metal and against controls.

The immunogen compound of the invention is typically composed of a biopolymer carrier, a spacer arm covalently bonded to the carrier and the heavy metal coordinated to the spacer arm. The spacer arm is preferably semi-rigid and has at least one heavy metal coordination site. This arrangement maintains the heavy metal in at least a partially exposed state and prevents substantially complete inclusion or chelation of the heavy metal by spacer arm and/or carrier. The biopolymer carrier may be a polysaccharide, a synthetic polyamide or preferably a protein. Preferred classes include blood or tissue sera proteins.

The spacer arm is generally no more than about 25 atoms in length. Preferably, the spacer arm is composed of one of three classes: an oligopeptide, an aliphatic compound or an aliphatic fragment. More preferably, the spacer arm is an oligopeptide. The first two classes are generally each substituted with no more than about 2 pendent Lewis acid or base groups, and a coupling group for forming a covalent bond with the protein carrier. The aliphatic fragment is substituted by a coupling group for forming a covalent bond with the protein carrier, and a carboxylic acid, hydroxyl, mercapto, amine or other group adapted for interacting with the heavy metal. For each class of spacer arm, the coupling group is an amine, carboxylic acid, aldehyde, hydroxyl or mercapto group.

A preferred spacer arm for metallic cations is an oligopeptide or aliphatic compound having no more than about 2 pendent Lewis base groups wherein the deformation of the electron shell of the Lewis base group is approximately of the same character as the deformation of the electron shell of the metallic cation. Especially preferred Lewis base groups for transition elements and the heavy metals are those containing sulfur, such as a sulfonate or thiol group. Especially preferred are spacer arms which include

glutathione and cysteine, mercapto ethanol amine, dithiothreitol, amines and peptides containing sulfur, and the like. Other preferred spacer arms include a carboxylic acid group or a phosphorus-containing Lewis base group such as a phosphonate anion.

The metallic cations are derived from metals such as period four transition metals, and period five, six and seven metals, transition elements and inner transition elements. The metallic cations of special mention as the heavy metal include those derived from zinc, lead, cadmium, bismuth, cobalt, arsenic, chromium, copper, nickel, strontium and mercury. Preferably, the metallic cations are lead cations, e.g., lead(II) cations.

The present invention also provides a method of forming a hybridoma which produces an antibody capable of immunoreacting with a lead cation and preferably with an exposed form of the lead cation. The method includes inoculating a mammal with a solution containing second antibodies which are capable of immunoreacting with the lead cation. Immune cells are isolated from the mammal and fused with an immortal mammal cell line from a second mammal. The resulting fused cell lines are selected for hybridomas which produce monoclonal antibodies capable of immunoreacting with a lead cation (Group 3 lead cation monoclonal antibodies; "Group 3 Pb mAbs").

Methods according to the invention utilize the metal binding polypeptide for detection, removal,

neutralization or addition of the heavy metal respectively in, from, within or to a solid, liquid or gaseous medium. These methods utilize features such as metal binding polypeptide immobilization, heavy metal immobilization, competitive binding, and means employing an oscillating probe, a micromagnetic probe and other physiochemical methods typically used to monitor antigen-antibody interactions.

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Methods for detection that are based upon heavy metal immobilization may indicate the presence of the heavy metal-metal binding polypeptide conjugate (e.g., a lead cation-Fab fragment conjugate) by known immunologic 5 assay techniques. In a first step, the heavy metal may be coordinated with an immobilized spacer arm. spacer arm can be any of the foregoing that will hold the heavy metal in at least a partially exposed state. It need not be the same spacer arm of the immunogen 10 compound used to develop the metal binding polypeptide. Non-immobilized materials are then removed from the mixture holding the immobilized spacer arm-heavy metal. Addition of the metal binding polypeptide (e.g., Fab fragment), removal of uncomplexed metal binding polypeptide and immunoassay complete the steps for this detection method.

Methods for detection that are based upon an immobilized metal binding polypeptide may utilize a radioactive version of the heavy metal or a similar tagged form thereof. Such tags include fluorescent, colorimetric and other spectrally active groups that can be coordinated or bonded to the heavy metal like the spacer arm. A preferred tag is a spacer arm containing a spectrally active group. First, the immobilized monoclonal antibody is saturated with the tagged heavy metal. After removal of the non-immobilized components, an aliquot of the unknown heavy metal is added. It displaces a portion of the bound, tagged heavy metal and measurement of that amount displaced will determine the concentration of unknown metal.

Methods for detection that are based upon an oscillating probe utilize either an immobilized spacer arm for the heavy metal or preferably immobilized metal binding polypeptide. This method measures the change in frequency of an oscillating surface as a function of the change in weight of that surface due to the binding of the non-immobilized heavy metal or metal binding

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polypeptides are immobilized on the surface of a high frequency oscillating probe. The probe is placed into a medium containing an unknown quantity of heavy metal. Binding of the heavy metal to the immobilized metal binding polypeptide will change the oscillation frequency of the probe. Hence, the degree of change will indicate the level of heavy metal present.

When the heavy metal is present as a metal cation in an aqueous medium, an especially preferred method for detection utilizes an oligopeptide having reactive group(s) capable of coordinating with the metal cation. The oligopeptide and the metal binding polypeptide specific for the metal cation unknown are added to the aqueous medium. The medium then is assayed for the presence of metal binding polypeptide cation conjugate. The interaction of the metal binding polypeptide with the metal cation is independent of the order of addition of the reactants and is independent of the identity of the oligopeptide.

In an especially preferred version of this method, a fixed support is utilized. Here, either the oligopeptide or the metal binding polypeptide is immobilized on the fixed support. The method is then conducted as related above.

The invention, in addition, contemplates methods for heavy metal removal from, heavy metal neutralization within or heavy metal addition to biological or inanimate systems. For all methods, an effective amount of the metal binding polypeptide is combined in some fashion with at least part of the system. Pursuant to the removal method, metal binding polypeptide-heavy metal conjugate is removed by separation means such as immunoprecipitation, immobilization, chromatography, filtration and the like. Pursuant to the neutralization method, the metal binding polypeptide-heavy metal conjugate remains in the system until it is removed by non-specific means. Pursuant to the addition method,

the metal binding polypeptide-heavy metal conjugate also remains in the system and the heavy metal is actively incorporated or otherwise used therein.

When the system participating in the foregoing methods is biological, the metal binding polypeptide may be combined with a pharmaceutically acceptable carrier. Preferably, the metal binding polypeptide will not of itself cause an undesirable immune response of the biological system. The biological systems contemplated according to the invention include unicellular organisms, multicellular simple organisms, cellular component systems, tissue cultures, plants and animals, including mammals.

The present invention also contemplates methods for removing heavy metallic cations or radioactive compounds from human fluids such as blood, serum or lymph by utilization of immobilized metal binding polypeptides.

An extracorporeal shunt placed in the patient permits removal of the body fluid and its reintroduction.

O Passing the body fluid extracorporeally through a bed of immobilized metal binding polypeptide accomplishes the desired removal.

The present invention also contemplates a kit for assaying the presence and quantity of heavy metal in a biological or inanimate system. The kit includes aliquots of metal binding polypeptides in the appropriate buffer, as well as a fixed support for absorption of the heavy metal, washing solutions, reagents such as enzyme substrates, and metal binding polypeptide specific antisera conjugated to a detectable substrate.

Brief Description of the Drawings

Figures 1A-1B depict the nucleotid and deduced
35 amino acid sequences for amino acids 1 through 113 of
the heavy chain variable regions of monoclonal
antibodies which immunoreact with a lead cation. The

numbering scheme is according to Kabat et al., Sequences of Proteins of Immunological Interest, vol. II. 5th edition, U.S. Department of Health and Human Services (1991) (hereinafter "Kabat"). Dashes indicate sequence identity with the 6B11 sequence; periods indicate gaps compared to the 1254 sequence.

Figure 2 depicts the nucleotide and deduced amino acid sequences for amino acids 5 through 108 of the light chain variable regions of monoclonal antibodies

which immunoreact with a lead cation. The numbering scheme is according to Kabat. Dashes indicate sequence identity with the 6B11 sequence.

Figures 3A-3B depict the nucleotide and deduced amino acid sequences for amino acids 1 through 113 of the heavy chain variable regions of monoclonal antibodies which immunoreact with a lead cation. The numbering scheme is according to Kabat. Periods indicate gaps.

Figure 4 depicts the nucleotide and deduced amino acid sequences for amino acids 1 through 107 of the light chain variable regions of monoclonal antibodies which immunoreact with a lead cation. The numbering scheme is according to Kabat.

Figure 5 depicts the nucleotide and deduced amino acid sequences for amino acids 1 through 113 of the heavy chain variable regions of monoclonal antibody 2B4 which immunoreacts with a lead cation. The numbering scheme is according to Kabat.

Figure 6 depicts the nucleotide and deduced amino acid sequences for amino acids 1 through 113 of the heavy chain variable regions of monoclonal antibody 2E7 which immunoreacts with a lead cation. The numbering scheme is according to Kabat.

Figure 7 shows the sensitivity of a set of Group 1 monoclonal antibodies (standardized to 50 μ g/mL in PBS) to lead (as Pb(NO₃)₂). The indicated amounts of lead were added to a microtiter plat coated with an amino

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acid polymer formed from glutamic acid, lysine, and tyrosine (hereinafter "EKY polymer") and analyzed by ELISA. The measured absorbances were corrected for the absorbance of an average of four control wells.

Figure 8 shows the sensitivity of a second set of Group 1 monoclonal antibodies (standardized to 50 μ g/mL in PBS) to lead (as Pb(NO₃)₂). The indicated amounts of lead were added to a microtiter plate coated with EKY polymer and analyzed by ELISA. The measured absorbances 10 were corrected for the absorbance of an average of four control wells.

Figure 9 shows the sensitivity of a set Group 3 monoclonal antibodies (standardized to 50 μ g/mL in PBS) to lead (as Pb(NO₃)₂). The indicated amounts of lead were added to a microtiter plate coated with EKY polymer and analyzed by ELISA. The measured absorbances were corrected for the absorbance of an average of four control wells.

Figure 10 shows the sensitivity of a second set of Group 3 monoclonal antibodies (standardized to 50 μ g/mL in PBS) to lead (as $Pb(NO_3)_2$). The indicated amounts of lead were added to a microtiter plate coated with EKY polymer and analyzed by ELISA. The measured absorbances were corrected for the absorbance of an average of four control wells.

Figure 11 shows the sensitivity of a third set of Group 3 lead cation monoclonal antibodies (standardized to 50 μ g/mL in PBS) to lead (as Pb(NO₃)₂). Monoclonal antibody 3A6, which is specific for the PR8 influenza 30 virus, was included as a control. The indicated amounts of lead were added to a microtiter plate coated with EKY polymer and analyzed by ELISA. The measured absorbances were corrected for the absorbance of an average of four control wells.

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Detailed Description of the Invention

Metal binding polypeptides of the present invention are key to the development of methods for detecting, adding, neutralizing or removing minute quantities of 5 heavy metals. Until the present invention, it was not possible to produce metal binding polypeptides which selectively bind with exposed heavy metal cations per se. The novel techniques for incorporating heavy metals into immunogen compounds and for administering these 10 immunogen compounds to immune cell hosts allow production of the desired, immunospecific monoclonal antibodies according to the invention. These methods are believed to constitute an advancement in the understanding of immunology.

Although not intended as a limitation of the invention, it is now believed that mammalian immunogenic reactivity can be elicited by heavy metals. While they are smaller than the commonly recognized epitope size of approximately 20-25 angstroms, the heavy metals nevertheless can bind.

Notwithstanding these beliefs, the invention contemplates metal binding polypeptides which selectively bind with a heavy metal, e.g. monoclonal antibodies to heavy metals. Hybridomas for the monoclonal antibodies and immunogen compounds for carrying the heavy metals and inducing immunogenicity are also included in the present invention. The metal binding polypeptides may include a monoclonal antibody, a recombinantly produced Fab fragment or a fusion protein. The fusion protein includes the heavy chain variable region of a monoclonal antibody which is capable of immunoreacting with a heavy metal, such as a lead cation. The invention also provides methods for the detection, addition, neutralization or removal of 35 heavy metals using the metal binding polypeptides.

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Monoclonal Antibodies The monoclonal antibodies of the invention are mammalian immunoglobulin proteins which have strong affinity constants for a specific heavy metal. Preferably, the monoclonal antibodies are from the IgG, IgA, IgM and IgE classes of antibodies, and more preferably, from the IgA, and IgM classes of antibodies. The heavy metal monoclonal antibodies are typically characterized by selective immunoreactivity with a particular heavy metal and a substantially lower immunoreactivity with other similarly structured heavy metals. Preferably, the monoclonal antibodies have an association constant for the selected heavy metal that is at least about 10,000 fold greater than the association constant for any similarly structured heavy metal. With respect to heavy metal cations, the especially preferred IgM and IgA classes of monoclonal antibodies of the present invention exhibit discriminatory dissociation constants of about 10-6M to about 10⁻¹²M. Examples include monoclonal antibodies of the IgM class which are produced by hybridomas 6B11, 1254, 7D10, 4E8, 8E7, 14F11 (ATCC NO. HB 11330), 10G5 and 14G11 and have a dissociation constant for lead cation of less than about 10-9M but do not bind cadmium, copper, zinc, nickel and cobalt cations to any appreciable extent. Other examples include monoclonal antibodies of the IgM and IgA classes which are produced by hybridomas derived from the spleen cells of a mammal injected with a solution of antibodies capable of immunoreacting with a lead cation. Suitable examples

immunoreacting with a lead cation. Suitable examples include monoclonal antibodies of the IgM class which are produced by hybridomas 13D10, 2E7, and 6F5. Monoclonal antibodies produced by hybridoma 11D11, 2B6, 13E8 and 8E7 are examples of antibodies of the IgA class which are capable immunoreacting with a lead cation.

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Immunogen Compounds

The immunogen compounds for generation of the specific immunogenicity of the monoclonal antibodies are based upon the hapten-carrier concept. The present 5 invention, however, broadens this concept so that the hapten is coordinated at the end of a spacer arm covalently bonded to the carrier. The spacer arm is adapted so as to be semi-rigid and to hold the heavy metal in an exposed position relative to the carrier. This arrangement is also adapted to maintain the heavy 10 metal in a substantially exposed and preferably, essentially completely exposed state. These factors combine substantially to avoid chelating, covering or inclusion of the heavy metal by the spacer arm and/or the carrier. 15

The spacer arm, as characterized above, may be an oligopeptide, an aliphatic compound, or an aliphatic fragment. In the latter two instances, the aliphatic compound or fragment may be covalently bonded to the carrier by means of a Schiff base reaction with an 20 aldehyde group, an amide formation reaction with an amine or carboxylic acid group using a peptide activator such as carbodiimide, acid chloride and the like, an ester formation reaction with a hydroxyl or carboxylic acid group using a Schotten Bauman reaction, or azide or 25 acid catalysis reaction, a sulfide reaction using a sulfide coupling agent, or other known coupling reactions for joining organic molecules to proteins. See for example Kabat, E.A., Structural Concepts In Immunology and Immunochemistry, 2nd Ed., Holt, Rinehart 30 and Winston, New York, 1976 (a review text of such methods) and Jaime Eyzaguirre, Chemical Modification of Enzymes: Active Site Studies, John Wiley & Sons (1982), the disclosures of which are incorporated herein by reference. The oligopeptide, aliphatic compound or 35 fragment will contain backbone groups which provide semi-rigidity to the spacer arm. Preferred groups for

developing this semi-rigidity include peptide bonds, olefin bonds, olefinic conjugated systems, ester groups and enone groups. Optionally, and especially where immunogenicity of the heavy metal appears difficult to generate, one or more aromatic rings can be incorporated into the spacer arm to stimulate the development of an immune response.

In general, the oligopeptide spacer arm has the following formula:

- X - (R) - Y

wherein X is a coupling group that will bond to the carrier, R is one or more amino acid residues and Y is the Lewis Acid or Base group(s) for heavy metal coordination.

In general, the aliphatic compound or fragment spacer arm has the following formula:

$$- X - (Q) - Z$$

wherein X is a coupling group that will bond to the carrier, Q is a semirigid aliphatic moiety containing 20 ester, amide, keto, olefin or aromatic groups and the like, and Z is a Lewis acid or Base group(s) for heavy metal coordination.

The oligopeptide or aliphatic compound is used as the spacer arm to coordinate a metal cation. In this instance, the pendent Lewis base groups will preferably be positioned at the spacer arm end remote from the carrier. These Lewis base groups function as the coordination site or sites for the metal cation. It is preferable that the deformability of the electron shells of the Lewis base groups and the metal cations be approximately similar. Accordingly, sulfur groups can serve as the Lewis base groups when the metal cations are transition metals or inner transition elements.

The carrier of the immunogen compound is a large 35 biopolymer that is known to participate in the development of hapten antigenicity. Blood serum proteins, amylopectins, polysaccharides, fetal serum

components, biologically acceptable natural and synthetic proteins and polyamides such as polyglycine can serve as the carriers. Preferred carriers include serum and tissue proteins. Examples are keyhole limpet 5 hemocyanin (KLH) and bovine serum albumin (BSA). Other examples include ovalbumin and chicken gamma globulin. These carriers have sites for coordinate bonding of the spacer arm. Such sites are preferably populated by amine groups, carboxylic acid groups, aldehyde groups and/or alcohol groups.

Production of Hybridomas

The production of hybridomas according to the invention generally follows the Kohler, Milstein technique cited supra. Many heavy metals, however, 15 toxify the mammalian system being used as a source of immune cells. This effect makes it important to determine the highest allowable dose of heavy metal and/or immunogen compound that can be used over a substantially long period of time without killing the 20 host. Pursuant to the Kohler, Milstein technique, immunization of the mammalian host may be accomplished within this dose parameter by subcutaneous or intraperitoneal injection of the immunogen compound in adjuvant. Administration is repeated periodically and preferably for at least four injections. Three days before the spleen is removed, a priming injection of immunogen compound is again administered.

After separation, the spleen cells are fused with immortal mammal cells such as mouse myeloma cells using 30 the techniques outlined by Kohler and Milstein. Polyethylene glycol (PEG) or electrical stimulation will initiate the fusions.

The fused cells are then cultured in cell wells according to culture techniques known in the art. Cellular secretions in the culture medium are tested

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after an appropriate time for the presence of the desired cellular products.

The induction of antibodies to heavy metals in a mammalian host may also be accomplished by subcutaneous or intraperitoneal injection of a lead cation monoclonal antibody solution in an adjuvant. These antibodies are typically derived from another animal of the same species. The spleen cells are then isolated and fused with immortal mammalian cells as described above. As before, the fused cells are cultured and the cellular secretions are examined for the presence of the desired antibodies ("Group 3 monoclonal antibodies").

The latter technique based on the induction of antibodies using a second lead cation antibody may be more efficient than the method based on the injection of 15 a Pb(II)/glutathione/carrier immunogen compound into a Table 1 herein shows the relative success rate for induction of lead cation antibodies using the two techniques. The results shown in Table 1 demonstrate that the rate of generating lead cation monoclonal 20 antibody producing hybridomas formation derived from injection of the 1254 mAb was greater than twenty fold more efficient than the method based on injection of the OVA/qlutathione/Pb(II) immunogen compound. single set of fusions of spleen cells from animals injected with the 1254 mAb produced more lead cation antibodies than the total number of lead cation antibodies produced by five sets of fusions of spleen cells induced with the immunogen compound.

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Selection Technique

The selection technique for identifying the appropriate monoclonal antibody is an important aspect for determining the immunospecificity desired according to the invention. The selection techniques according to the invention call for determining the binding affinity of the hybridoma cellular products against the heavy

metal and against cross-reactive controls. In particular, hybridoma culture fluid is tested in screening assays against the heavy metal, the carrier, the carrier-spacer arm product and the immunogen compound as well as optionally against the spacer armheavy metal coordinate. Screening assays can be performed by immunoenzymatic-assay, immunofluorescence, radioimmunoassay, immunoprecipitative assay or inhibition of biological activity.

The hybridoma cultures selected will exhibit strong binding characteristics to the heavy metal (and immunogen compound) and will not bind with the spacer arm-carrier product or with the carrier itself.

Following the identification of cell cultures

15 producing the desired monoclonal antibodies, subcloning
to refine the selected culture can be performed. These
techniques are known to those skilled in the art. See
for example Goding, James Goding, Monoclonal Antibodies:
Principles and Practice, 2nd Edition, Academic Press,

- 20 San Diego, CA 1986, the disclosure of which is incorporated herein by reference. Briefly, the appropriately selected cell culture is separated into one cell units which are then recultured. The subclone cultures are then again tested for specific
- immunoreactivity, lack of cross-reactivity and the amount of monoclonal antibody secreted. Those subcultures exhibiting the highest amounts of secreted monoclonal antibody are chosen for subsequent pilot development.

Following the foregoing techniques, a number of hybridomas producing monoclonal antibodies to lead cations have been developed. Perpetual cell lines, designated 6B11, 1254 (8.15), 7D10, 4E8, 8E7, 17E5, 14F11, 10G5, 14G11 and 3H7, produce monoclonal antibodies to lead cations. The group of cell lines were developed by inoculation of mice with an

OVA/glutathione/Pb(II) conjugate. The sensitivity of

monoclonal antibodies produced by these hybridoma cell lines is shown in Figures 7 and 8. The sensitivity of mAb 3A6 (a monoclonal antibody specific for the PR8 influenza virus), which was included as a control, as anticipated, showed no senisitivity to Pb(II) (see Figure 8). Monoclonal antibody 4A10 was derived from the spleen cells of a mouse injected with a mercury cation. The monoclonal antibodies produced by hybridoma 4A10 cross react with Pb(II) cations as well as immunoreacting with a mercury cation.

Using similar fusion and selection techniques, a number of hybridomas producing monoclonal antibodies to lead cations were developed by inoculation of mice with different lead cation monclonal antibodies ("Group 3 lead cation monoclonal antibodies"). Perpetual cell 15 lines, designated 2B4, 2B6, 3B6, 13B7, 13B11, 16B11, 6C2, 14C6, 19C6, 11D11, 13D10, 2E7, 4E4, 12E5, 13E8, 18E7, 6F5, 1G4, 1G6, 6G2, and 13G9 were developed using this latter technique. The sensitivity of monoclonal 20 antibodies produced by these hybridoma cell lines is shown in Figures 9-11. The set of Group 3 lead cation monoclonal antibodies exhibited lead sensitivity comparable to the "parent" Group 1 antibody, mAb 1254, and to other Group 1 lead cation monoclonal antibodies 25 generated by injection of the OVA/glutathione/lead(II) immunogen compound.

The immunogenic host for these hybridomas was the BALB/c mouse and the fusion partner was chosen from the mouse myeloma cell lines P3X63-Ag8.653 or SP2/0.

30 Immunizations to induce Group 1 lead cation antibodies were accomplished with the immunogen compound formed from ovalbumin, glutathione and lead(II) cation functioning as the heavy metal in complete Freund's adjuvant. Immunizations to induce Group 3 lead cation antibodies were accomplished with the lead cation monoclonal antibody designated 1254 ("mAb 1254") in an adjuvant. All of the cell lines mentioned above are

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maintained in culture medium and in frozen medium at liquid nitrogen temperature.

PCR Amplification

PCR amplification of Fd and κ regions from the spleen messenger RNA (mRNA) of a mouse immunized with BSA-glutathione-lead cation may be performed as described by Sastry et al., Proc. Natl. Acad. Sci. U.S.A., 86, 5728 (1989). The PCR amplification is performed with cDNA obtained by the reverse transcription of the mRNA with a primer specific for amplification of heavy chain sequences or light chain sequences.

The PCR amplification of mRNA isolated from spleen cells or hybridomas with oligonucleotides that incorporate restriction sites into the ends of the amplified product may be used to clone and express heavy chain sequences (e.g., the amplification of the Fd fragment) and κ light chain sequences from mouse spleen cells. The oligonucleotide primers, which are analogous to those that have been successfully used for amplification of V_R sequences (see Sastry et al., Proc. Natl. Acad. Sci U.S.A., 86, 5728 (1989)), may be used for these amplifications. Restriction endonuclease recognition sequences are typically incorporated into these primers to allow for the cloning of the amplified fragment into a λ phage vector in a predetermined reading frame for expression.

30 <u>Expression of Fab Fragments on Phage Coat</u>

Phage assembly proceeds via an extrusion-like process through the bacterial membrane. Filamentous phage M13 has a 406-residue minor phage coat protein (cpIII) which is expressed before extrusion and which accumulates on the inner membrane facing into the periplasm of <u>E. coli</u>. The two functional properties of cpIII, infectivity and normal (nonpolyphage)

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morphogenesis, have been assigned to roughly the first and second half of the gene. The N-terminal domain of cpIII binds to the F' pili, allowing for infection of E. coli, whereas the membrane-bound C-terminal domain, P198-S406, serves the morphogenic role of capping the trailing end of the filament according to the vectorial polymerization model.

A phagemid vector may be constructed to fuse the antibody Fd chain with the C-terminal domain of cpIII 10 (see Barbas et al., Proc. Natl. Acad. Sci. USA, 88, 7978 (1991)). A flexible five-amino acid tether (GGGGS), which lacks an ordered secondary structure, may be juxtaposed between the expressed Fab and cpIII domains to minimize interaction. The phagemid vector may also 15 be constructed to include a nucleotide coding for the light chain of a Fab fragment. The cpIII/Fd fragment fusion protein and the light chain protein may be placed under control of separate lac promoter/operator sequences and directed to the periplasmic space by pelB 20 leader sequences for functional assembly on the membrane. Inclusion of the phage F1 intergenic region in the vector allows for packaging of single-stranded phagemid with the aid of helper phage. The use of helper phage superinfection may result in expression of 25 two forms of cpIII. Consequently, normal phage morphogenesis may be perturbed by competition between the cpIII/Fd fragment fusion protein and the native cpIII of the helper phage for incorporation into the The resulting packaged phagemid may carry 30 native cpIII, which is necessary for infection, and the fusion protein including the Fab fragment, which may be displayed for interaction with an antigen and used for selection. Fusion at the C-terminal domain of cpIII is necessitated by the phagemid approach because fusion 35 with the infective N-terminal domain would render the host cell resistant to infection. The result is a phage displaying antibody combining sites ("Phabs").

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antibody combining sites, such as Fab fragments, are displayed on the phage coat. This technique may be used to produce Phabs which display recombinantly produced Fab fragments (e.g., recombinantly produced Fab fragments that immunoreact with a lead cation) on the phage coat of a filamentous phage such as M13.

A phagemid vector (pComb 3) which allows the display of antibody Fab fragments on the surface of filamentous phage, has been described (see Barbas et 10 al., Proc. Natl. Acad. Sci. USA, 88, 7978 (1991)). XhoI and SpeI sites for cloning PCR-amplified heavy-chain Fd sequences are included in pComb 3. SacI and XbaI sites are also provided for cloning PCR-amplified antibody light chains. These cloning sites are compatible with 15 known mouse and human PCR primers (see, e.g., Huse et al., Science, 246, 1275-1281 (1989)). The nucleotide sequences of the pelB leader sequences are recruited from the λ HC2 and λ LC2 constructs described in Huse et al, ibid, with reading frames maintained. Digestion of pComb 3 encoding a selected Fab with SpeI and NheI permits the removal of the gene III fragment. Because SpeI and NheI produce compatible cohesive ends, the digested vector may also be religated to yield a phagemid that produces soluble Fab.

Phabs may be produced by overnight infection of phagemid containing cells (e.g., infected <u>E. coli</u> XL-1 Blue) yielding typical titers of 10¹¹ cfu/mL. By using phagemids encoding different antibiotic resistances, ratios of clonally distinct phage may easily be

determined by titering on selective plates. In single-pass enrichment experiments, clonally mixed phage may be incubated with an antigen-coated plate. Nonspecific phage will be removed by washing, and bound phage may then be eluted with acid and isolated.

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Methods of Application

According to the invention, the metal binding polypeptide can be used to advantage for detection, neutralization, addition or removal of heavy metals from These methods apply to 5 biological or inanimate systems. qualitative and quantitative analyses of minute concentrations of toxic metal cations, in aqueous liquid systems, in biological or environmental systems or in such compositions as perfumes, cosmetics, 10 pharmaceuticals, health care products, skin treatment products, pesticides, herbicides, solvents used in the production of semi-conductor and integrated circuit components and production materials for electronic

components. In each application, the presence of minute 15 quantities of metallic cations could constitute deleterious contaminants. Their ready and early detection will avoid later production or regulatory setbacks.

Alternatively, the presence of minute quantities of heavy metals in certain instances may be desirable. example, the presence of inorganic moieties in such mixtures as doping materials for semi-conductors and integrated circuits contributes to the properties of the product. Quality control of the presence and concentration of these heavy metals is essential for the functioning of the product. The detection methods of the invention enable ready and early measurement of the presence of such heavy metals and avoid later production or regulatory difficulties.

Heavy metals in biological or inanimate systems can also be removed by methods according to the invention. In the main, immobilization of the metal binding polypeptides on a solid support followed by its mixture with the materials of the biological or inanimate system 35 will remove the heavy metals. In this instance, the immobilization of the monoclonal antibodies can be accomplished by techniques known to those of skill in

the art. <u>See</u>, for example, <u>Affinity Chromatography</u>,
C.R. Fowe & P.D.G. Sean, John Wiley & Sons, London 1974,
the disclosure of which is incorporated herein by
reference. Removal is accomplished by passing a fluid
mixture of the system ingredients suspected of having
the heavy metals over the immobilized metal binding
polypeptides. Of course, the metal binding polypeptides
are designed to be specific for the heavy metal sought
to be removed.

An advantage of this method is the removal of undesirable heavy metals in the presence of similarly structured desirable metal species. For example, whole blood from a patient suffering from lead poisoning can be removed from the patient, optionally filtered to return the cellular blood components to the patient, and the serum or blood passed over immobilized metal binding polypeptides specific for the lead. The purified serum or blood can then be returned to the patient. The lead will be removed but other blood serum components such as zinc, calcium and the like will not.

Specific Applications

A particular application of the present invention contemplates a method for the production of monoclonal antibodies specific for the lead cation or another 25 toxic, heavy metal cation. In accordance with this method, the heavy metal cation in question is combined into an immunogen compound as described above and suspended in an aqueous medium. A preferred protein 30 carrier for the immunogen compound in this instance is an albumin, e.g., ovalbumin. keyhole limpet hemocyanin is also a preferred carrier. The preferred spacer arm in this instance is an oligopeptide which has sulfonate or carboxylate groups capable of coordinating with the 35 heavy metal cation. The suspension of immunogen compound is used to immunize a host mammal such as a mouse following the techniques outlined above.

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laboratory strain of mouse designated BALB/c is particularly preferred.

Spleen cells of the immunized host are collected and converted into a suspension. These spleen cells are 5 fused with immortal cells as described above. Preferably, myeloma cells of the same animal species as the immunized host are used as the fusion partner. Typically, a cell fusion promoter such as polyethylene glycol is employed to cause formation of the hybridoma cells. The hybridoma cells are diluted and cultured in 10 a medium which does not allow for the growth of unfused cells.

The monoclonal antibodies produced and secreted by the hybridomas are thereafter assayed for the ability to 15 bind immunologically with the heavy metal cations used for immunization. The antibodies are further selected for lack of cross-reactivity with carrier, with carrierspacer arm and with similar metallic cations. preferred assay method in this context is an enzyme-linked immunosorbent assay.

The resulting monoclonal antibodies are specific for heavy metal cations and exhibit strong immunoreaction to the heavy metal cations in the presence of spacer arm or the spacer arm-carrier composition. Preferred monoclonal antibodies are selectively immunoreactive with lead cations.

The present invention also provides methods and kits for detecting the presence of a toxic heavy metal cation such as a lead cation. The methods and kits include a metal binding polypeptide which selectively binds the heavy metal.

According to an embodiment of a method for detecting the presence of a specific heavy metal cation, an immobilized coordinating compound is combined with the unknown mixture containing the toxic heavy metal The heavy metal cation complexes with the coordinating compound and is immobilized thereto.

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Removal of the non-immobilized components leaves the immobilized toxic heavy metal cation. Addition of the metal binding polypeptide specific for the targeted heavy metal cation forms an immobilized cation-metal binding polypeptide conjugate. Its presence and concentration can be assayed by an ELISA technique or other tagging or visualization technique known to those of skill in the art. In this process, of course, non-immobilized metal binding polypeptide is removed before the assay is conducted.

A kit for quantitatively measuring the presence of a heavy metal cation is a further aspect of the invention. The kit includes a metal binding polypeptide specific for the toxic metal cation in question. metal binding polypeptide is preferably metered into several aliquots of varying, known concentration. kit may also include the immobilized coordination compound, preferably, attached to a solid support such as the well of a microtiter plate or a chromatographic 20 material. The kit also typically includes a visualization or tagging assay material for determination of the presence of the metal binding polypeptide-heavy metal cation conjugate. If desired, a meter or other device for detecting and signaling the level of visual or other reading from the assay may also be included.

The invention will be further characterized by the following examples. These examples are not meant to limit the scope of the invention which has been fully set forth in the foregoing description. Variation within the concepts of the invention are apparent to those skilled in the art.

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Example 1 Lead Cation Monoclonal Antibodies

A. General Procedures

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1. <u>Generation of Hybridomas via Injection of OVA/glutathione/Pb(II)</u>

Hybridoma antibodies were produced from the spleen cells of BALB/c mouse that had received multiple

10 injections of lead(II) cations reacted with glutathione to produce a lead cation coordinate covalent compound, which was covalently bound to ovalbumin

("OVA/glutathione/Pb(II) antigen"). Glutathione is a three amino acid residue peptide having one reactive sulfhydryl group which forms a coordinate bond with lead cations. The OVA/glutathione/Pb(II) antigen in complete Freund's adjuvant was utilized to assist in the elicitation of an immune response in the host animal.

Of hybridomas isolated, some were determined to be producing monoclonal antibody specific for glutathione. In addition, other hybridomas (6B11, 1254, 8E7, 10G5 and 14F11) were producing monoclonal antibodies that were strongly positive against glutathione-lead cation but negative against glutathione without lead cation. These antibodies were subcloned by the process of limiting dilution for further characterization.

ELISA Analysis for Hybridoma Antibodies Immunoreactive With Glutathione-Lead Cations

Microtiter plates (EIA/RIA grade) were treated with BSA-glutathione, blocked with 1% polyvinyl alcohol in phosphate-buffered saline ("PBS") and used for the ELISA. One hundred microliters of lead nitrate (100 ppm in 100 mMolar Hepes pH 7.2) was added to the wells and incubated for 30 minutes. The plates were washed three times with PBS containing 0.1% Triton X-100, and then the hybridoma culture supernatant was added for 30 minutes at room temperature. After washing the wells, goat anti-mouse antibodies conjugated to horseradish peroxidase were added. Following incubation for 30

minutes at room temperature, the plates were washed with PBS containing 0.1% Triton X-100, and 100 ul of ABTS peroxidase substrate was added to each well. After 15 min at room temperature the absorbance of each well was read at 405 nanometers. In addition, the frozen hybridoma samples have been thawed from liquid nitrogen and assayed for persistence of antibody secretion after thawing.

10 2. <u>Determination of Lead-Cation Specific</u> Monoclonal Antibodies

The monoclonal antibodies may be assayed with various other metals for inhibition of binding of the monoclonal antibodies to lead cations. The cationic

15 metals assayed may include the ions of zinc, copper, cadmium, nickel, and arsenic. For instance, the inhibition of the binding of a monoclonal antibody which immunoreacts with immobilized glutathione-lead cations by various concentrations of divalent cations may be

20 examined. Metal ions at the indicated concentrations are incubated with culture fluid from the antibody in an ELISA plate. The absorbance at 405 nm may be determined for each sample, and the percent inhibition of each metal ion concentration determined by the formula given above.

Further analysis of the monoclonal antibodies can establish if the antibodies are specific for the lead cations per se and that glutathione is not needed for the monoclonal antibodies to react with and bind to the lead cations. The monoclonal antibodies may also be assayed against BSA-glutathione, BSA-glutathione-lead cations, and BSA-lead cations. Comparison with a negative control consisting of a monoclonal antibody specific for an unrelated antigen permits a determination of whether the monoclonal antibodies bind to lead cation in the absence of glutathione.

PBS containing metal ions at the indicated concentrations may be added to microtiter wells to which

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BSA-glutathione has been absorbed. After incubation at room temperature for 30 minutes, the plates are washed to remove unbound metals, and the plates are used for a standard ELISA to detect lead cations.

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B. Particular Preparations

1. Linkage of Lead Cations to Protein Carriers

To prepare antigen for injection and immunoassay, 132 mg Pb(NO_3)₂ (400 μ mol), 61 mg glutathione (200 μ mol) and 54 mg NaCl were dissolved in 10 mL of water. After 10 cold ethanol (30mL) was added, the resulting mixture was incubated for 30 minutes at 0°C. The reaction mixture was centrifuged at 10,000 g for 30 minutes, and the pellet was washed with 30 mL of cold ethanol. pellet was dissolved in 200 mL of 40% dimethylformamide pH 4.8, containing 200 mg of 1-ethyl-3-(3dimethylaminopropyl)-carbodiimide, and l g of either bovine serum albumin or OVA was added to the solution. The reaction mixture was stirred at room temperature 20 overnight. The mixture was then centrifuged as above, resuspended in PBS, and dialyzed overnight at 4°C against 4 liters of PBS.

2. <u>Immunization of BALB/c Mice</u>

25 Multiple injections of the antigen prepared with 10 ug of protein per injection were made into BALB/c mice. Lead cation-glutathione-OVA emulsified in Freund's adjuvant was employed as the antigen. Complete adjuvant was used for the first two injections, while incomplete adjuvant was used for all subsequent injections. After the fourth injection, a drop of blood from the tail of each mouse was collected separately in 0.5 mL of PBS, and each sample was assayed by ELISA for the presence of antigen-specific antibody. The mice used for hybridoma production received an intraperitoneal injection consisting of 10 ug of antigen in PBS 3-4 days before cell fusion.

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3. <u>Hybridoma Production</u>

The spleen was removed aseptically from a mouse. and the cells were isolated by placing the spleen in 5 mL of sterile PBS and teasing it with two sterile, 5 18-gauge hypodermic needles. The cell suspension was added to an empty sterile, conical, 15-mL centrifuge tube and tissue fragments were allowed to settle for 1-2 minutes. The cells still in suspension were placed in a tube similar to that above and centrifuged at 300 g for 10 minutes at room temperature. The cells were then washed 3 times by centrifugation in serum-free DMEM (Dulbecco's modified Eagle's medium). Spleen cells were co-pelleted with P3X63-Ag8.653 myeloma cells at a ratio of 4 spleen cells to 1 myeloma cell. The supernatant fluid was removed, and the pellet was suspended in 1 mL of 35% polyethylene glycol for 1 minute. The polyethylene glycol was gradually diluted by addition of increasing amounts of serum-free DMEM over a period of 15 minutes. The cells are then suspended in HAT medium 20 (Monoclonal Antibodies, Kennett, McKean, Backitt, eds., Plenum Press (1981)) at a concentration of 2 X 10⁵ myeloma cells per mL, and 100 μ L of the suspension were added to each well of 5 separate 96-well microtiter plates. The plates were incubated in 10% CO, at 37°C for 25 one week. At that time, half of the culture fluid was withdrawn from each well and replaced by 2 drops of fresh HT medium (HAT medium without aminopterin), and the plates were incubated as above for another week. Then, approximately 100 μL of culture fluid was taken 30 from each well containing macroscopically visible cell growth, and the ELISA technique described infra was used for identification of those culture fluids containing lead cation-specific antibodies.

35 4. Enzyme-Linked Immunosorbent Assay (ELISA)

Polyvinyl chloride microtiter assay plates may be coated with antigen by addition of 50 μ l of lead

cation-glutathione-BSA or glutathione-BSA at a concentration of 5 ug/mL in PBS to each well of the plate. The plates are allowed to incubate at room temperature overnight to allow the antigen to dry on the plate. Next day the plates are blocked by addition of 200 μL of 1% polyvinyl alcohol ("PVA") in PBS to each well; the addition of the PVA blocks the remaining protein-binding sites. The plates are incubated for 30 min at room temperature, then washed 3 times with ELISA wash (PBS with 0.1% of Triton X-100), 3 times with milliQ water. A 100 ppm solution of lead in 100mM Hepes at pH 7 is then added to the wells (100 μL/well and allowed to incubate for 30-60 minutes at room temperature.

15 Fifty microliters of culture fluid to be assayed for the presence of antigen-specific antibody may be added to the appropriate well, and the plates are incubated at room temperature for 2 hours. The plates are again washed 3 times with ELISA wash, and 50 µL of 20 goat anti-mouse serum (Cooper Biomedical) diluted 1:1000 in 2% BSA in PBS are added to each well. After incubation and washing as above, 50 µL of rabbit anti-goat serum conjugated to alkaline phosphatase (Sigma) diluted 1:1000 in 50 mM Tris-HCl, pH 8.0, 25 containing 1 mM MgCl₂, 5% BSA and 0.04% NaN₃, are added to each well. After being incubated and washed as above, 150 μ L of phosphatase substrate (0.4 mM dinitrophenyl phosphate in 1 M diethanolamine, pH 9.8, containing 25 mM MgCl₂) are added to each well.

The enzyme catalyzed conversion of dinitrophenol phosphate to dinitrophenol is typically allowed to proceed at room temperature for 30-60 minutes. The absorbance of each well at 405 nm (dinitrophenol) may be measured with a UV/Vis spectrometer.

The use of other enzymes as sensors is also possible provided that such enzymes can be linked to an appropriate antibody, and catalyze a reaction which

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produces a color change. For example, beta galactosidase, urease, or horseradish peroxidase could be utilized in this context.

5 5. <u>Binding of Lead Cations to Immobilized Coordinating Spacer Arms</u>

One hundred microliters of BSA-glutathione at a concentration of 5 ug/mL were added to the wells of a microtiter plate and allowed to dry overnight. The 10 plates were then blocked with PVA as above. One hundred microliters of PBS containing a known concentration of lead cations were added to triplicate wells on the plate, which were then incubated at room temperature for 30 minutes. After this incubation period the plates are washed with ELISA wash to remove unbound metal ions and then used in the standard ELISA to measure reactivity with a lead cation-specific antibody.

6. Assay of Lead Cation-Specific Antibodies Against BSA Glutathione, BSA Glutathione-Lead and BSA-Lead Lead cation specific antibodies secreted from hybridomas such as 6B11, 1254, 8E7, 10G5 and 14F11 were assayed against BSA-glutathione and BSA-glutathione-lead cation. The results established that all five antibodies immunoreacted with BSA-glutathione that had bound lead, and did not immunoreact with BSA-glutathione

30 Nucleotides Coding for Heavy Chain Fd
Fragments and Light Chains from Lead
Cation Monoclonal Antibodies

lacking lead.

Synthesis of Nucleotides Encoding the Heavy and Light
Chain Variable Regions of the Lead-Cation Antibodies

RNA was isolated from hybridoma cells with
guanidine isothiocyanate (Evans et al., BioTechniques,
8, 357 (1990)), and enriched for poly(A) + RNA by
passage over a poly(dT)-cellulose column (Aviv et al.,

40 Proc. Natl. Acad. Sci. USA, 69, 1408 (1972)). First-

strand cDNA synthesis was catalyzed by MuLv reverse transcriptase with a Promega RiboClone kit, according to the manufacturer's directions. The primers used for cDNA synthesis were complementary to the 5' end of the $C_{\rm H}1$ domain of the heavy chain expressed by the hybridoma of interest, or to the 5' end of the $C_{\rm K}$ domain. The following primers were used for cDNA synthesis:

KAPPA Primer - Light Chain
51-GAAGATCTAGACTTACTATGCAGCATCAGC-31 (SEQ ID

10 NO:41)

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MU primer - Heavy Chain

5¹-AGGAGACTAGTGGTTACTAATTTGGGAAGGACTG-3¹ (SEQ ID NO:42)

Amplification of Antibody Variable Regions by Polymerase Chain Reaction

The primer used for cDNA synthesis of the variable region of a particular antibody polypeptide chain was also used for PCR amplification of that variable region, in conjunction with an appropriate V-region primer as described in Huse et al., Science, 246, 1275 (1989).

The PCR was performed as described in Sastry et al., Proc. Natl. Acad. Sci. USA, 86, 5728 (1989).

25 <u>Sequence Determination of Nucleotides Encoding the Heavy</u> <u>and Light Chain Variable Regions of the Lead Cation</u> Antibodies

The PCR amplified nucleotide sequences encoding the heavy and light chain variable regions of the lead cation antibodies were cloned into Bluescript (Stratagene, La Jolla, CA). The sequences of these nucleotides were determined by the dideoxy chain termination method (Sanger et al., Proc. Natl. Acad.
Sci. USA, 74, 5463 (1977)). The sequences of at least three PCR products for each heavy and light chain were determined to allow detection of incorporation errors by Taq polymerase. The nucleotide and deduced amino acid sequences of the heavy and light chain variable regions

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of the lead-specific antibodies are shown in Figures 1A-1B and 2.

Figures 1A-1B depict the nucleotide and deduced amino acid sequences for the heavy chain variable regions of a number of monoclonal antibodies that immunoreact with a lead cation. The following sequences are shown:

the heavy chain variable region nucleotide acid sequence and deduced amino acid sequence for monoclonal antibody 6B11 (SEQ ID NO:1);

the heavy chain variable region nucleotide sequence and deduced amino acid sequence for monoclonal antibody 1254 (SEQ ID NO:3);

the heavy chain variable region nucleotide sequence and deduced amino acid sequence for monoclonal antibody 8E7 (SEQ ID NO:5);

the heavy chain variable region nucleotide sequence and deduced amino acid sequence for monoclonal antibody 10G5 (SEQ ID NO:7); and

the heavy chain variable region nucleotide sequence and deduced amino acid sequence for monoclonal antibody 14F11 (SEQ ID NO: 9).

Figure 2 depicts the nucleotide and deduced amino acid sequences for the light chain variable regions of a number of monoclonal antibodies which immunoreact with a lead cation. The following sequences are shown:

the light chain variable region nucleotide sequence and deduced amino acid sequence for monoclonal antibody 6B11 (SEQ ID NO: 11);

the light chain variable region nucleotide sequence and deduced amino acid sequence for monoclonal antibody 1254 (SEQ ID NO:13); and

the light chain variable region nucleotide sequence and deduced amino acid sequence for monoclonal antibody 14F11 (SEQ ID NO:15).

Figures 3A-3B depict the nucleotide and deduced amino acid sequences for the heavy chain variable

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regions of a number of monoclonal antibodies that immunoreact with a lead cation. The following sequences are shown:

the heavy chain variable region nucleotide acid sequence and deduced amino acid sequence for monoclonal antibody 13D10 (SEQ ID NO:17);

the heavy chain variable region nucleotide sequence and deduced amino acid sequence for monoclonal antibody 11D11 (SEQ ID NO:19);

the heavy chain variable region nucleotide sequence and deduced amino acid sequence for monoclonal antibody 14G11 (SEQ ID NO:21);

the heavy chain variable region nucleotide sequence and deduced amino acid sequence for monoclonal antibody 6F5 (SEQ:ID NO:23);

the heavy chain variable region nucleotide sequence and deduced amino acid sequence for monoclonal antibody 7D10 (SEQ ID NO:25); and

the heavy chain variable region nucleotide sequence and deduced amino acid sequence for monoclonal antibody 4E8 (SEQ ID NO:27).

Figure 4 depicts the nucleotide and deduced amino acid sequences for the light chain variable regions of a number of monoclonal antibodies which immunoreact with a lead cation. The following sequences are shown:

the light chain variable region nucleotide sequence and deduced amino acid sequence for monoclonal antibody 10G5 (SEQ ID NO:29);

the light chain variable region nucleotide sequence and deduced amino acid sequence for monoclonal antibody 2E7 (SEQ ID NO:31); and

the light chain variable region nucleotide sequence and deduced amino acid sequence for monoclonal antibody 7D10 (SEQ ID NO: 33).

Figure 5 depicts the nucleotide and deduced amino acid sequences for the heavy chain variable regions of

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the monoclonal antibody 2B4 that immunoreacts with a lead cation. The following sequences are shown:

the heavy chain variable region nucleotide sequence and deduced amino acid sequence for monoclonal antibody 2B4 (SEQ ID NO:35).

Figure 6 depicts the nucleotide and deduced amino acid sequences for the heavy chain variable regions of the monoclonal antibody 2E7 that immunoreacts with a lead cation. The following sequences are shown:

the heavy chain variable region nucleotide sequence and deduced amino acid sequence for monoclonal antibody 2E7 (SEQ ID NO: 37).

As can be seen in the above-described amino acid sequences, (see, for example, figures 2 and 4) most of the light chains of the lead-reactive monoclonal antibodies are members of the V_k32 light chain family, having a motif of four carboxylic acid-containing residues, e.g., glutamic acid and/or aspartic acid residues, in a stretch of five amino acids in the CDR1. Results of molecular modeling indicate that one or more of the carboxyl groups of these residues are positioned at the appropriate distance to form coordinate

As described in the Examples, regardless of the specific antigen used for induction of lead-specific antibodies (e.g., anti-lead antibody or lead-containing complex), the majority of the resulting lead-reactive monoclonal antibodies contained light chains of the $V_{\rm k}32$ family.

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1.

Example 3 Expression of Fusion Proteins Including Heavy Chain Fd Fragments and Light Chains from Lead

Vector Construction

interractions with lead.

The pelB leader sequences and cloning sites for the heavy-chain fragment and light chain may be derived from phagemids excised from λ Hc2 and λ Lc2 vectors as

Cation Monoclonal Antibodies

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described in Huse, et al., Science, 246, 1275-1281 (1989). The sequences may be modified to remove a redundant SacI site from Hc2 phagemid and a SpeI site from the Lc2 phagemid. The combinatorial phagemid vector pComb is constructed from these two modified phagemids by restricting each with ScaI and EcoRI and combining them in a ligation reaction. Recombinants are screened for the presence of two NotI sites yielding the combinatorial vector pComb. The tether sequence GGGGS and gIII fragment (gene coding for coat protein III of filamentous phage M13 (see Barbas, et al., Proc. Natl. Acad. Sci. USA, 88, 7978 (1991)) from SpeI to NheI are the product of PCR of M13mp18 (Yanisch-Perron, et al., Gene, 33, 103-119 (1985)) using the oligonucleotides 5'-GAGACGACTAGTGGTGGCGGTGGCTCTCCATTCGTTTGTGAATATCAA-3' (SEQ ID NO: 43) and 5'-TTACTAGCTAGCATAATAACGGAATACCCAAAAGAACTGG-3' (SEO ID as reported in Barbas, et al., Proc. Natl. Acad. Sci. USA, 88, 7978 (1991).

20 The lacZ promoter, operator, and Cap-binding site controlling light chain expression are the product of PCR with M13mp18 using oligonucleotides 5'-TATGCTAGCTAGTAACACGACAGGTTTCCCGACTGG-3' (SEO ID NO:45) and 5'-AGCTTTGAATTCGTGAAATTGTTATCCGCT-3' (SEO ID 25 NO:46) as reported in Barbas et al., ibid. The PCR fragments encoding the gIII fragment and lacZ promoter are spliced by PCR overlap extension (see Horton et al., Gene, 77, 61-68 (1989)). The resulting product is digested with SpeI and EcoRI and ligated into the 30 corresponding sites of pComb to yield pComb 3'. Finally, pComb 3' is digested with XhoI and SpeI and ligated with the corresponding 51-base-pair (bp) stuffer from pBluescript (see Short, et al., Nucleic Acids Res., 16, pp. 7583-7600 (1988)) (Stratagene) to yield pComb 3, 35 an ampicillin-resistant phagemid.

2. <u>Expression of Nucleotides on M13 Phage Coat</u> <u>Phage Production</u>

A pComb 3 phagemid including a recombinantly produced Fab fragment that immunoreacts with a lead 5 cation may be transformed into E. coli XL1-Blue cells. The transformed E. coli XL1-Blue cells may be grown in super broth medium (SB; 30 g of tryptone, 20 g of yeast extract, 10 g of Mops per liter, pH 7) at 37°C supplemented with tetracycline at 10 μ g/mi and 10 carbenicillin at 50 μ g/mL or chloramphenicol at 30 $\mu g/mL$. Cultures are grown to an OD_{600} of 0.4 and infected with VCSM13 helper phage (phage to cell ratio, 20:1) and grown an additional hour. After one hour kanamycin is added (70 μ g/mL), and the culture is 15 incubated overnight at 30°C. Phage may be isolated from liquid culture by polyethylene glycol 8000 and NaCl precipitation as described in Cwirla, et al., Proc. Natl. Acad. Sci. USA, 87, pp. 6378-6382 (1990). Phage pellets may be resuspended in phosphate-buffered saline (10 mM phosphate, pH 7.2, 150 mM NaCl) and stored at 20 -20° C.

3. Single-Pass Enrichment Experiments

Phage expressing lead cation binding Fab fragments on their surface may be enriched by a modification of 25 the panning procedure described by ParmLey, et al., Gene, 73, pp. 305-318 (1988). A single well of a microtiter plate (Costar 3690) is coated overnight at 4° C with 25 μ L of BSA-glutathione-lead cation at 2 mg/mL in 0.1 M bicarbonate, pH 8.5. The well is washed once with water and blocked by filling the well with Blotto (1% (wt/vol) PVA in phosphate-buffered saline) and incubating the plate at 37°C for one hour. Blocking solution is shaken out, and 50 μ l of clonally mixed phage (typically 1011 colony-forming units (cfu)) is added, and the plate is incubated for an additional 2 hr at 37°C. Phage are removed, and the well is washed once with distilled water. The well is washed 10 times with TBS/Tween solution (50mM Tris-HCl, pH 7.5, 150 mM NaCl,

0.05% Tween 20) over a period of one hour at room temperature. The well is washed once more with distilled water, and adherent phage are eluted by adding 50 μL of elution buffer (0.1 M HCl, adjusted to pH 2.2
5 with glycine), and incubation at room temperature for 10 min. The eluate is removed and neutralized with 3 μL of 2 M Tris base. The initial phage input ratio may be determined by titering on selected plates. The final phage output ratio may be determined by infecting 1 mL
10 of logarithmic phase XL1-Blue cells with the neutralized eluate for 15 min at room temperature and plating equal aliquots on selective carbenicillin and chloramphenicol plates.

4. Lead Cation-ELISA for Phabs

Equal plaque forming units from Phabs obtained from cultures of E. coli XL1-Blue cell may be transformed with a phagemid including a recombinantly produced Fab fragment that immunoreacts with a lead cation incubated at 37°C for two hours on BSA-Glutathione ELISA plates

with or without lead nitrate. A rabbit anti-M13 antiserum is typically used as a second antibody followed by affinity-purified goat-antirabbit serum conjugated with peroxidase. 2,2'-Azino-Di-[3-ethylbenzthiazoline sulfonate] (ABTS) may be used as peroxidase substrate. The results are typically expressed as absorbance at 405 nanometers.

5. pComb3 Phagemid Expressing 1254 Light Chain
Using the methods described above, a pComb3

30 phagemid expressing only the light chain of monoclonal antibody 1254, without any accompanying heavy chain, was constructed. Two copies of the 1254 light chain sequence were cloned into the phagemid. Using the ELISA assay described above, the phagemid reacted with lead,

35 demonstrating that the light chain of the antibodies is sufficient for lead binding.

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Example 4 Group 3 Lead Cation Monoclonal Antibodies

Generation of Hybridomas via Injection of Lead 1. Cation Monoclonal Antibody 1254

5 Group 3 antibodies were produced by injecting mice with the Group 1 lead cation monoclonal antibody, mAb 1254 (mAb 8.15). The mice were injected biweekly with 50 μ g of antibody emulsified in Freunds complete adjuvant and bled seven days after each injection. 10 mice were bled by nicking the tail and collecting 50 ul of blood, which was diluted to 500 μ l in phosphate buffered saline (PBS). The serum was tested using the same ELISA procedure as for screening the fusions (described below). The spleen cells of the mice were then used for production of hybridoma monoclonal 15 antibodies (mAb's) as described below.

Fusion Procedure for Hybridoma Production

A mouse was sacrificed by cervical dislocation, and its spleen was removed aseptically. A single cell suspension was prepared from the spleen, and the cells were washed three times in serum-free Dulbecco's modified Eagle's medium (DMEM) by centrifugation at 400x g for 10 minutes, with 10 ml of DMEM per wash. number of viable spleen cells was determined by trypan 25 blue exclusion with a hemacytometer. The spleen cells were then mixed with SP2/0 myeloma cells at a ration of 4 spleen cells:1 myeloma cell. The SP2/0 cells had previously been washed three time with serum-free DMEM as described above for the spleen cells.

The cell mixture was centrifuged as above, and the cell pellet was resuspended in 500 μ l of 50% polyethylene glycol (PEG 1500, Sigma Chemical Co., St. Louis, MO), which was added over a one-minute period. The cell suspension was stirred for an addition minute, followed by addition of 10 ml of serum-free DMEM over the next two minutes. After stirring for an additional minute, the cells were centrifuged at 400x g for 10 minutes, and the pellet was resuspended in HAT medium to

a final concentration of 1 x 10⁷ spleen cells/ml. Two hundred microliters of the cell suspension was added to each well of the appropriate number of 96-well microtiter plates, and the fusion was incubated at 37°C in an atmosphere of 10% CO₂, 90% air. All reagents and supplies used in this procedure were sterile and at room temperature.

Culture media

All culture media was filtered through a 0.2 μ m 10 pore size filter (Nalgene).

A. Complete media

-Add the following to 500 milliliters of DMEM (Gibco):

-10% of complement free (56 C for 1 hour)

bovine calf serum (Hyclone)

-45 millimolar of sodium pyruvate (Sigma)

-200 millimolar of L-glutamine (Sigma)

-0.1% of gentamicin (Gibco)

B. HT media

20 -Add the following to 500 milliliters of DMEM:

-20% of complement free (56°C for 1 hour)

fetal clone (Hyclone)

-45 millimolar of sodium pyruvate (Sigma)

-200 millimolar of L-glutamine (Sigma)

-10 millimolar of hypoxanthine (Sigma)

-1.5 millimolar thymidine (Sigma)

-100 millimolar oxaloacetate (Sigma)

-0.1% of gentamicin (Gibco)

C. HAT media

-Add the following to 500 milliliters of DMEM:

-20% of complement free (56 C for 1 hour)

fetal clone (Hyclone)

-40 micromolar aminopterin (Sigma)

-45 millimolar of sodium pyruvate (Sigma)

35 -200 millimolar of L-glutamine (Sigma)

-10 millimolar of hypoxanthine (Sigma)

-1.5 millimolar thymidine (Sigma)

-100 millimolar oxaloacetate (Sigma)
-20 I.U./milliliter of bovine insulin (Sigma)
-0.1% of gentamicin (Gibco)

An amino acid polymer consisting of glutamic acid, lysine, and tyrosine at a ratio of 6:3:1 respectively ("EKY polymer"; Sigma Chemical Co.) was prepared as a 50 μg/ml solution in PBS. One hundred microliters of the EKY polymer solution was added to each well of a high-binding polystyrene plate (Corning Costar, Cambridge, MA). The plate was incubated at room temperature for thirty minutes, then rinsed three times with distilled water. One hundred microliters of 1% polyvinyl alcohol (w/v) in PBS was added to each well, and the plate was

A Standard Reference Material (SRM) 3128 (National Institute of Standards and Technology, Gaithersburg, MD) 20 consisting of 10 mg/ml lead nitrate in 10% HNO₃ was diluted-tenfold in 1.0 M HEPES, pH 9.5. This solution was diluted to a final concentration of 100 μg/ml in 0.1 M HEPES, pH 8.0. One hundred microliters of the resulting solution was added to each well of the plates treated with the EKY polymer. The plates were incubated for 30 minutes and washed three times with distilled water before being used for ELISA.

incubated at room temperature for sixty minutes, after

which it was rinsed as above.

One hundred microliters of hybridoma culture fluid to be assayed for the presence of lead cation monoclonal antibodies was added to the well of an antigen-coated microliter plate. The assay plate was incubated for 30 minutes at room temperature. The plate was then rinsed three times with ELISA wash (0.1% Triton X 100 in PBS) and three times with distilled water. One hundred microliters of a 1:1000 dilution of goat anti-mouse IgA, IgG, IgM (Kirkegaard and Perry Laboratories, Gaithersburg, MD) conjugated to horseradish peroxidase

was added to each well. The plate was incubated at room temperature for thirty minutes, then rinsed as described above. After one hundred microliters of ABTS peroxidase substrate (Kirkegaard and Perry Laboratories) were added to each well and the plate was incubated at room temperature for fifteen minutes. The absorbance of each well at 405 nanometers was then measured on a Bio Tek Model EL-311 Plate Reader (Fisher, Pittsburgh, PA). The measured absorbances were compared to the absorbance of an average of four control wells. A hybridoma was considered positive if the absorbance of an assay well containing lead was twice that of a control well to which no lead had been added.

Positive hybridomas were subcloned by limiting
dilution in 96-well microtiter plates in medium
consisting of 50% HT medium and 50% complete DMEM
conditioned by buffalo rat liver cells. Buffalo rat
liver cells are known to secrete several somatomedin
growth factors into the medium during growth. The
somatomedin growth factors enhance the growth of cells
in cloning situations.

3. <u>Precipitation of Monoclonal Antibodies from Ascites</u> <u>Fluid</u>

25 Ascites fluid was centrifuged at 300g for 10 minutes at 4°C to remove any particulate matter. The supernant fluid was decanted into a centrifuge tube and chilled in an ice bath for fifteen minutes. An ammonium sulfate solution was prepared by mixing 9 parts of a 30 saturated solution of ammonium sulfate with 1 part distilled water. The resulting ammonium sulfate solution was added dropwise to an equal volume of ascites fluid while stirring gently. The mixture was stirred for an addition 4 hours at 4°C. The precipitate 35 was pelleted by centrifugation at 3000 g for 30 minutes at 4°C. The pellet was resuspended in and dialyzed against PBS at 4°C.

The volume of the dialysate was measured, and two volumes of 60 millimolar sodium acetate, pH 4.0, was added to bring the final pH to 4.8. Caprylic acid (Sigma Chemical Co.) was added dropwise at a ratio of 0.4 milliliters per ten milliliters of the original volume of ascites fluid. The mixture was stirred at 4°C for four hours and centrifuged as above. The pellet was resuspended in PBS and dialyzed against PBS with 0.05% Triton X-100 for 24 hours then against PBS for 24 hours.

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4. <u>Measurement of the Sensitivity of the Lead-Specific</u> <u>Monoclonal Antibodies by ELISA</u>

ELISA was carried out essentially as described in subsection 2 above except that the lead nitrate solution was diluted to final concentrations of 100, 50, 10, 5, or 1 μ g/ml using 0.1 M HEPES, pH 8.0. One hundred microliters of each of these solutions was then added to the plate coated with the polyamino acid mixture. Caprylic acid-precipitated mAb was adjusted to a concentration of 50 μ g/ml, and 100 μ l was added to the appropriate wells of the microliter plate. The plate was incubated for 30 minutes at room temperature, and the ELISA was continued as described above.

25 All publications and patent applications in this specification are indicative of the level of ordinary skill in the art to which this invention pertains. All publications and patent applications are herein incorporated by reference to the same extent as if each 30 individual publication or patent application was specifically and individually indicated by reference.

The invention has been described with reference to various specific and preferred embodiments and techniques. However, it should be understood that many variations and modifications may be made while remaining within the spirit and scope of the invention.

SEQUENCE ID NO. DESIGNATIONS

SEQ ID #	ANTIBODY NAME	DESCRIPTION	SEQUENCE TYPE
1	6B11	Heavy Chain	Nucleic Acid
2	6B11	Heavy Chain	Amino Acid
3	1254	Heavy Chain	Nucleic Acid
4	1254	Heavy Chain	Amino Acid
5	8E7	Heavy Chain	Nucleic Acid
6	8E7	Heavy Chain	Amino Acid
7	10G5	Heavy Chain	Nucleic Acid
8	10G5	Heavy Chain	Amino Acid
9	14F11	Heavy Chain	Nucleic Acid
10	14F11	Heavy Chain	Amino Acid
11	6B11	Light Chain	Nucleic Acid
12	6B11	Light Chain	Amino Acid
13	1254	Light Chain	Nucleic Acid
14	1254	Light Chain	Amino Acid
15	14F11	Light Chain	Nucleic Acid
16	14F11	Light Chain	Amino Acid
17	13D10	Heavy Chain	Nucleic Acid
18	13D10	Heavy Chain	Amino Acid
19	11D11	Heavy Chain	Nucleic Acid
20	11D11	Heavy Chain	Amino Acid
21	14G11	Heavy Chain	Nucleic Acid
22	14G11	Heavy Chain	Amino Acid
23	6F5	Heavy Chain	Nucleic Acid
24	6 F 5	Heavy Chain	Amino Acid
25	7D10	Heavy Chain	Nucleic Acid
26	7D10	Heavy Chain	Amino Acid
27	4E8	Heavy Chain	Nucleic Acid
28	4E8	Heavy Chain	Amino Acid
29	10G5	Light Chain	Nucleic Acid
30	10G5	Light Chain	Amino Acid
31	2E7	Light Chain	Nucleic Acid
32	2E7	Light Chain	Amino Acid
33	7D10	Light Chain	Nucleic Acid
34	7D10	Light Chain	Amino Acid
35	2B4	Heavy Chain	Nucleic Acid
36	2B4	Heavy Chain	Amino Acid
37	2E7	Heavy Chain	Nucleic Acid
38	2E7	Heavy Chain	Amino Acid
39	6F5	Light Chain	Nucleic Acid
40	6F5	Light Chain	Amino Acid
41	KAPPA Primer	Light Chain	Nucleic Acid
42	MU Primer	Heavy Chain	Nucleic Acid
43	n.a. primer	·····	Nucleic Acid
44	n.a. primer	Ticht Obsis	Nucleic Acid
45	n.a. primer	Light Chain	Nucleic Acid
46	n.a. primer	Light Chain	Nucleic Acid

Table 1. Comparision of the Fusion Efficiencies in Group 1 versus Group 3 Fusions

Group/fusion	# Cells Fused	# Positives	# Positives/107cells fused
1A	19.0 x 10 ⁷	5	0.26
1B	18.6 x 10 ⁷	1	0.05
1C	11.2 x 10 ⁷	1	0.09
1D	18.7×10^7	1	0.05
1E	12.2 x 10 ⁷	l	0.08
Average	15.9 x 10 ⁷	2.8	0.18
3	17.7 x 10 ⁷	66	3.73
Control	14.5×10^7	0	0.0

Table 2. Class Analysis of Group 1 Lead Antibodies

#	Group	Hybridoma	Heavy Chain	Light Chain
1	1	8.15	μ	κ
2	1 .	4A10	μ	κ
3	1	3H7	μ	κ
4	1	4E8	μ	κ .
5	1	6 B 11	μ	κ
6	1	7D10	μ	κ
7	1	8E7	μ .	· K
8	1	10G5	μ	κ
9	1	14F11	μ	κ
10	1	14 G 11	μ	κ
11	1	17E5	μ	ĸ

Table 3. Class Analysis of Group 3 Lead-Specific Antibodies

#	Group	Clone	Heavy Chain	Light Chain
1	3	1G4	μ	κ
2	3	1 G 6	μ	κ
3	3	2B4	μ	ĸ.
4	3	2B6	α	κ
5	3	2E7	μ	μ
6	3	3B6	μ	κ
7	3	4E4	μ	κ
8	3	6C2	μ	κ
9	3	6F5	μ	· K
10	3	6G2	μ	κ
11	3	11D11	α	λ
12	3	12E5	μ	κ
13	3	13B7	μ	κ
14	3	13 B 11	μ	κ
15	3	13 D 10	μ	κ
16	3	13E8	α	κ
17	3	13 G 9	μ	κ
18	3	14C6	μ	κ
19	3	16B11	μ	κ

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Table 3. Class Analysis of Group 3 Lead-Specific Antibodies (continued)

20	3	18E7	α	κ
21	3	19C6	μ	κ

SEQUENCE LISTING

(1) GENERAL INFORMATION

- (i) APPLICANT:
 - (A) NAME: BioNebraska, Inc.
 - (B) STREET: 3820 Northwest 46th Street
 - (C) CITY: Lincoln (D) STATE: Nebraska

 - (E) COUNTRY: United States of America
 - (F) POSTAL CODE (ZIP): 68524
- (ii) TITLE OF THE INVENTION: LEAD BINDING POLYPEPTIDES AND NUCLEOTIDES CODING THEREFORE
- (iii) NUMBER OF SEQUENCES: 46
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: Merchant, Gould, Smith, Edell, Welter & Schmidt
 - (B) STREET: 3100 Norwest Center, 90 South Seventh St
 - (C) CITY: Minneapolis
 - (D) STATE: MN
 - (E) COUNTRY: USA
 - (F) ZIP: 55402
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Diskette
 - (B) COMPUTER: IBM Compatible
 - (C) OPERATING SYSTEM: DOS
 - (D) SOFTWARE: FastSEQ Version 1.5
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: PCT/US96/09258
 - (B) FILING DATE: 05-JUN-1996
 - (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: 08/541,373
 - (B) FILING DATE: 10-OCT-1995
- (vii) PRIOR APPLICATION DATA:
 (A) APPLICATION NUMBER: 08/462,798
 - (B) FILING DATE: 05-JUN-1995
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Kettelberger, Denise M
 - (B) REGISTRATION NUMBER: 33,924
 - (C) REFERENCE/DOCKET NUMBER: 8648.49WOI2
- (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: 612/371-5268
 - (B) TELEFAX: 612/332-9081
 - (C) TELEX:

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 354 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:
 (ix) FEATURE:
- - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...354
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

GAG Glu 1	GTT Val	CAG Gln	CTG Leu	CAG Gln 5	CAG Gln	TCT Ser	GGA Gly	CCT Pro	GAG Glu 10	CTG Leu	GTG Val	AGG Arg	CCT Pro	GGA Gly 15	CCT Pro	48	
	GTG Val															96	
	ATG Met															144	
	ATG Met 50															192	
	GAC Asp															240	
	CAA Gln															288	
	AGA Arg															336	
	GTC Val															354	Ļ

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 118 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Glu Val Gln Leu Gln Gln Ser Gly Pro Glu Leu Val Arg Pro Gly Pro 10 Ser Val Lys Leu Ser Cys Lys Ala Ser Gly Tyr Ser Phe Thr Ser Tyr 25 Trp Met Asn Trp Val Lys Gln Arg Pro Gly Arg Gly Leu Glu Trp Ile 40 Gly Met Ile His Pro Ser Asp Ser Glu Thr Arg Leu Asn Gln Lys Phe 55 Lys Asp Lys Ala Thr Leu Thr Val Asp Lys Ser Ser Ser Thr Ala Tyr 70 75 Met Gln Leu Ser Ser Pro Thr Ser Glu Asp Ser Ala Val Tyr Tyr Cys 90 85 Ala Arg Arg Gly Asn Ser Ala Trp Phe Ala Tyr Trp Gly Gln Gly Thr 100 105 Leu Val Thr Val Ser Ala 115

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 366 base pairs

 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISKNSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...366
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

		CAG Gln 5							 48
		TCC Ser						_	 96
_		GTA Val				_			 144
	 	 CCT Pro	 	 			 		 192
		 ACA Thr	 	 			 		 24 0
		AGA Arg 85					_		 288
		GGG Gly							336
		CTG Leu							366

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 122 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Gln Val Gln Leu Gln Gln Ser Gly Ala Gly Leu Val Lys Pro Gly Ala 10 Ser Val Lys Leu Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Glu Tyr 25 Ile Ile His Trp Val Lys Gln Arg Ser Gly Gln Gly Leu Glu Trp Ile Gly Trp Phe Tyr Pro Gly Ser Gly Ser Ile Lys Tyr Asn Glu Lys Phe 50 55 60 Lys Asp Lys Ala Thr Leu Thr Ala Asp Lys Ser Ser Ser Thr Val Tyr 65 70 75 80 Met Glu Leu Ser Arg Leu Thr Ser Glu Asp Ser Ala Val Tyr Phe Cys 85 90 Ala Arg His Glu Gly Tyr Gly Asn Tyr Val Ala Trp Phe Ala Tyr Trp 100 105 Gly Gln Gly Thr Leu Val Thr Val Ser Ala 115 120

- (2) INFORMATION FOR SEQ ID NO:5:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 357 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...357
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

GTC Val			 		 			 	48
GTG Val	 		 	-,				 	96
ATG Met		_						 	144
AGG Arg 50									192
GAC Asp			 						240
CAA Gln									288
 AGA Arg	 		 					 	336
 TCA Ser	 		 						357

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 119 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Glu Val Gln Leu Gln Gln Ser Gly Ala Glu Leu Val Lys Pro Gly Ala 10 15 Pro Val Lys Leu Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Ser Tyr 20 25 30 Trp Met Asn Trp Val Lys Gln Arg Pro Gly Arg Gly Leu Glu Trp Ile 35 40 45 Gly Arg Ile Asp Pro Ser Asp Ser Glu Thr His Tyr Asn Gln Lys Phe 60 50 55 Lys Asp Lys Ala Thr Leu Thr Val Asp Lys Ser Ser Ser Thr Ala Tyr 75 80 65 70 Ile Gln Leu Ser Ser Leu Thr Ser Glu Asp Ser Ala Val Tyr Tyr Cys 95 90 Ala Arg His His Tyr Gly Tyr Tyr Ala Met Asp Tyr Trp Gly Gln Gly 110 105 100 Thr Ser Val Thr Val Ser Ser 115

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 354 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE: (ix) FEATURE:
- - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...354
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

GTC Val			 				 	48
GTG Val							 	96
ATG Met								144
ATG Met 50	 	 	 	 		 	 	192
GAC Asp	 	 	 				 	2 4 0
CAA Gln								288
AGA Arg								336
 GTC Val	 	 						354

- (2) INFORMATION FOR SEQ ID NO:8:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 118 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Glu Val Gln Leu Gln Gln Ser Gly Ala Glu Leu Val Arg Pro Gly Ala Ser Val Lys Leu Ser Cys Lys Ala Ser Gly Tyr Ser Phe Thr Ser Tyr 25 Trp Met Asn Trp Val Lys Gln Arg Pro Gly Gln Gly Leu Glu Trp Ile Gly Met Ile His Pro Ser Asp Ser Glu Thr Arg Leu Asn Gln Lys Phe Lys Asp Lys Ala Thr Leu Thr Val Asp Lys Ser Ser Ser Thr Ala Tyr 65 80 70 75 Met Gln Leu Ser Ser Pro Thr Ser Glu Asp Ser Ala Val Tyr Tyr Cys 95 90 85 Ala Arg Arg Arg Asp Tyr Asp Pro Phe Ala Tyr Trp Gly Gln Gly Thr 105 100 Leu Val Thr Val Ser Ala 115

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 357 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single

 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence (B) LOCATION: 1...357

 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

GAG Glu 1	GTC Val	CAG Gln	CTG Leu	CAG Gln 5	CAG Gln	TCT Ser	GGA Gly	GCT Ala	GAG Glu 10	CTG Leu	GTA Val	AGG Arg	CCT Pro	GGG Gly 15	ACT Thr	48
							GCT Ala									96
							AGG Arg 40									144
							GGT Gly									192
							GCA Ala									240
	_						TCT Ser					-				288
							TGG Trp									336
			ACC Thr													357

- (2) INFORMATION FOR SEQ ID NO:10:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 119 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Glu Val Gln Leu Gln Gln Ser Gly Ala Glu Leu Val Arg Pro Gly Thr 15 Ser Val Lys Val Ser Cys Lys Ala Ser Gly Tyr Ala Phe Thr Asn Tyr 20 Leu Ile Glu Trp Val Lys Gln Arg Pro Gly Gln Gly Leu Glu Trp Ile 35 40 45 Gly Val Ile Asn Pro Gly Ser Gly Gly Thr Asn Tyr Asn Glu Lys Phe 50 55 60 Lys Gly Lys Ala Thr Leu Thr Ala Asp Lys Ser Ser Ser Thr Ala Tyr 70 75 Met Gln Leu Ser Ser Leu Thr Ser Asp Asp Ser Ala Val Tyr Phe Cys 90 Ala Arg Ser Gly Tyr Gly His Trp Tyr Phe Asp Val Trp Gly Ala Gly 100 105 Thr Thr Val Thr Val Ser Ser 115

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 312 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...312 (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

					TCC Ser												48
					AGC Ser											;	96
CAG Gln	CAG Gln	AAG Lys 35	CCA Pro	GGG Gly	GAA Glu	CCT Pro	CCT Pro 40	AAA Lys	CTC Leu	CTT Leu	ATT Ile	TCA Ser 45	GAA Glu	GGC Gly	AAT Asn	1	44
					GTC Val											19	92
					ACA Thr 70											24	10
					CAA Gln											28	38
					ATA Ile											3	112

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 104 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Thr Gln Ser Pro Ser Ser Leu Ser Met Ala Ile Gly Glu Lys Val Thr 10 Ile Arg Cys Ile Thr Ser Thr Asp Ile Asp Asp Asp Met Asn Trp Tyr Gln Gln Lys Pro Gly Glu Pro Pro Lys Leu Leu Ile Ser Glu Gly Asn 35 40 Thr Leu Arg Pro Gly Val Pro Ser Arg Phe Ser Ser Ser Gly Tyr Gly 55 60 Thr Asp Phe Val Phe Thr Ile Glu Asn Met Leu Ser Glu Asp Val Ala 65 70 75 Asp Tyr Tyr Cys Leu Gln Ser Asp Asn Leu Pro Leu Thr Phe Gly Gly 90 Gly Thr Lys Leu Glu Ile Lys Arg 100

(2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 312 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE: (ix) FEATURE:
- - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...312
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

	CAG Gln																48
ATC Ile	AGA Arg	TGC Cys	ATA Ile 20	ACC Thr	AGC Ser	ACT Thr	GAT Asp	ATT Ile 25	GAT Asp	GAT Asp	GAT Asp	ATG Met	AAC Asn 30	TGG Trp	TAC Tyr		96
CAG Gln	CAG Gln	AAG Lys 35	CCA Pro	GGG Gly	GAA Glu	CCT Pro	CCT Pro 40	aaa Lys	CTC Leu	CTT Leu	ATT Ile	TCA Ser 45	GAA Glu	GGC Gly	AAT Asn	1	144
ACT Thr	CTT Leu 50	CGT Arg	CCT Pro	GGA Gly	GTC Val	CCA Pro 55	TCC Ser	CGA Arg	TTC Phe	TCC Ser	AGC Ser 60	AGT Ser	GGC Gly	TAT Tyr	GGT Gly	1	L92
ACA Thr 65	GAT Asp	TTT Phe	GTT Val	TTT Phe	ACA Thr 70	ATT Ile	GAA Glu	AAC Asn	ATG Met	CTC Leu 75	TCA Ser	GAA Glu	GAT Asp	GTT Val	GCA Ala 80	2	240
GAT Asp	TAC Tyr	TAC Tyr	TGT Cys	TTG Leu 85	CAA Gln	AGT Ser	GAT Asp	AAC Asn	TTG Leu 90	CCT Pro	CTC Leu	ACG Thr	TTC Phe	GGT Gly 95	GCT Ala	2	888
	ACC Thr																312

- (2) INFORMATION FOR SEQ ID NO:14:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 104 amino acids

 - (B) TYPE: amino acid (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Thr Gln Ser Pro Ser Ser Leu Ser Met Ala Ile Gly Glu Lys Val Thr 10 Ile Arg Cys Ile Thr Ser Thr Asp Ile Asp Asp Asp Met Asn Trp Tyr 20 25 30 Gln Gln Lys Pro Gly Glu Pro Pro Lys Leu Leu Ile Ser Glu Gly Asn Thr Leu Arg Pro Gly Val Pro Ser Arg Phe Ser Ser Ser Gly Tyr Gly Thr Asp Phe Val Phe Thr Ile Glu Asn Met Leu Ser Glu Asp Val Ala 75 65 70 Asp Tyr Tyr Cys Leu Gln Ser Asp Asn Leu Pro Leu Thr Phe Gly Ala 90 85 95 Gly Thr Lys Leu Glu Leu Lys Gly 100

(2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 312 base pairs
 (B) TYPE: nucleic acid

 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (iii) HYPOTHETICAL: NO

- (iv) ANTISENSE: NO (v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence (B) LOCATION: 1...312

 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

ACC Thr 1	CAG Gln	TCT Ser	CCA Pro	GCA Ala 5	TCC Ser	CTG Leu	TCC Ser	ATG Met	GCT Ala 10	ATA Ile	GGA Gly	GAA Glu	AAA Lys	GTC Val 15	ACC Thr	48
ATC Ile	AGA Arg	TGC Cys	ATA Ile 20	ACC Thr	AGC Ser	ACT Thr	GAT Asp	ATT Ile 25	GAT Asp	GAT Asp	GAT Asp	ATG Met	AAC Asn 30	TGG Trp	TAC Tyr	96
CAG Gln	CAG Gln	AAG Lys 35	CCA Pro	GGG Gly	GAA Glu	CCT Pro	CCT Pro 40	AAG Lys	CTC Leu	CTT Leu	ATT Ile	TCA Ser 45	GAA Glu	GGC Gly	AAT Asn	144
					GTC Val											192
					ACA Thr 70											240
					CAA Gln											288
					CTG Leu											312

- (2) INFORMATION FOR SEQ ID NO:16:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 104 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Thr Gln Ser Pro Ala Ser Leu Ser Met Ala Ile Gly Glu Lys Val Thr Ile Arg Cys Ile Thr Ser Thr Asp Ile Asp Asp Met Asn Trp Tyr 20 25 Gln Gln Lys Pro Gly Glu Pro Pro Lys Leu Leu Ile Ser Glu Gly Asn 35 40 45 Thr Leu Arg Pro Gly Val Pro Ser Arg Phe Ser Ser Ser Gly Tyr Gly 55 60 Thr Asp Phe Val Phe Thr Ile Glu Asn Met Leu Ser Glu Asp Val Ala 75 70 Asp Tyr Tyr Cys Leu Gln Ser Asp Asn Leu Pro Leu Thr Phe Gly Ala 90 Gly Thr Lys Leu Glu Leu Lys Arg 100

(2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 357 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE: (ix) FEATURE:
- - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...357
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

		 	 	 		GTG Val			48
						TCA Ser			96
		 	 	 	 	AAC Asn			144
						TAC Tyr 60			192
						AAG Lys			240
						GCC Ala			288
						TAC Tyr			336
	ACT Thr	 			,				357

(2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 119 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

Asp Val Gln Leu Gln Glu Ser Gly Pro Gly Leu Val Lys Pro Ser Gln 10 Ser Leu Ser Leu Thr Cys Thr Val Thr Gly Tyr Ser Ile Thr Ser Asp 30 Tyr Ala Trp Asn Trp Ile Arg Gln Phe Pro Gly Asn Lys Leu Glu Trp 35 40 Met Gly Tyr Ile Ser Tyr Ser Gly Ser Thr Ser Tyr Asn Pro Ser Leu 50 55 60 Lys Ser Arg Ile Ser Ile Thr Arg Asp Thr Ser Lys Asn Gln Phe Phe 65 70 75 Leu Gln Leu Asn Ser Val Thr Thr Glu Asp Thr Ala Thr Tyr Tyr Cys 85 90 95 Ala Arg Cys Gly Asn Tyr Pro Trp Tyr Phe Asp Tyr Trp Gly Gln Gly 100 105 Thr Thr Leu Thr Val Ser Ser 115

(2) INFORMATION FOR SEQ ID NO:19:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 357 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence (B) LOCATION: 1...357

 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

CAG Gln 1	GTT Val	CAG Gln	CTG Leu	CAG Gln 5	CAG Gln	TCT Ser	GGA Gly	GTT Val	GAG Glu 10	CTG Leu	ATG Met	AAG Lys	CCT Pro	GGG Gly 15	GCC Ala	48	
TCA Ser	GTG Val	AAG Lys	ATA Ile 20	TCC Ser	TGC Cys	AAG Lys	GCT Ala	ACT Thr 25	GGC Gly	TAC Tyr	ACA Thr	TTC Phe	AGT Ser 30	AGC Ser	TAC Tyr	96	
TGG Trp	ATA Ile	GAG Glu 35	TGG Trp	GTA Val	AAG Lys	CAG Gln	AGG Arg 40	CCT Pro	GGA Gly	CAT His	GGC Gly	CTT Leu 45	GAG Glu	TGG Trp	ATT Ile	144	
											TAC Tyr 60					192	
AAG Lys 65	GGC Gly	AAG Lys	GCC Ala	ACA Thr	TTC Phe 70	ACT Thr	GCA Ala	GAT Asp	ACA Thr	TCC Ser 75	TCC Ser	AAC Asn	ACA Thr	GCC Ala	TAC Tyr 80	240	
ATG Met	CAA Gln	GTC Val	AGC Ser	AGC Ser 85	CTG Leu	ACA Thr	TCT Ser	GAG Glu	GAC Asp 90	TCT Ser	GCC Ala	GTC Val	TAT Tyr	TAC Tyr 95	TGT Cys	288	
GCA Ala	AGG Arg	ATC Ile	TAC Tyr 100	TAT Tyr	GGT Gly	CAC His	TTG Leu	TGG Trp 105	TTT Phe	GCT Ala	TAC Tyr	TGG Trp	GGC Gly 110	CAA Gln	GGG Gly	336	
				GTC Val											٠	35	7

WO 96/39518 PCT/US96/09250

- (2) INFORMATION FOR SEQ ID NO:20:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 119 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

Gln Val Gln Leu Gln Gln Ser Gly Val Glu Leu Met Lys Pro Gly Ala Ser Val Lys Ile Ser Cys Lys Ala Thr Gly Tyr Thr Phe Ser Ser Tyr 20 25 Trp Ile Glu Trp Val Lys Gln Arg Pro Gly His Gly Leu Glu Trp Ile 40 Gly Glu Ile Leu Pro Gly Ser Gly Ser Thr Asn Tyr Asn Glu Lys Phe 60 Lys Gly Lys Ala Thr Phe Thr Ala Asp Thr Ser Ser Asn Thr Ala Tyr 75 70 Met Gln Val Ser Ser Leu Thr Ser Glu Asp Ser Ala Val Tyr Tyr Cys 90 85 Ala Arg Ile Tyr Tyr Gly His Leu Trp Phe Ala Tyr Trp Gly Gln Gly 105 100 Thr Leu Val Thr Val Ser Ala 115

(2) INFORMATION FOR SEQ ID NO:21:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 354 base pairs

 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE: (ix) FEATURE:
- - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...354
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

					CTT Leu				48
	_	 	 	 	 TTC Phe	_	 		96
		 	 	 	 CAG Gln				144
			 	 	GAA Glu				192
		 	 	 	 TCC Ser 75		 	 :	240
					ACT Thr			;	288
					TAC Tyr			;	336
GTC Val		 							354

(2) INFORMATION FOR SEQ ID NO:22:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 118 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

Glu Val Gln Leu Gln Gln Ser Gly Ala Glu Leu Val Arg Ser Gly Ala Ser Val Lys Leu Ser Cys Thr Ala Ser Gly Phe Asn Ile Lys Asp Tyr 20 Tyr Met His Trp Val Lys Gln Arg Pro Glu Gln Gly Leu Glu Trp Ile Gly Trp Ile Asp Pro Glu Asn Gly Asp Thr Glu Tyr Asp Pro Lys Phe 55 60 50 Gln Gly Lys Ala Thr Met Thr Ala Asp Thr Ser Ser Asn Ile Ala Tyr 65 70 75 80 Leu Gln Leu Ser Ser Leu Thr Ser Glu Asp Thr Ala Val Tyr Tyr Cys 90 85 95 Asn Pro Tyr Gly Tyr Asp Asp Ala Met Asp Tyr Trp Gly Gln Gly Thr 100 105 110 Ser Val Thr Val Ser Ser 115

(2) INFORMATION FOR SEQ ID NO:23:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 330 base pairs
 (B) TYPE: nucleic acid

 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:
 (ix) FEATURE:
- - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...330
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

	TCT Ser															48
TGT Cys	GCA Ala	GCC Ala	TCT Ser 20	GGA Gly	TTC Phe	ACC Thr	TTC Phe	AAT Asn 25	ACC Thr	TAC Tyr	GCC Ala	ATG Met	AAC Asn 30	TGG Trp	GTC Val	96
CGC Arg	CAG Gln	GCC Ala 35	CCA Pro	GGA Gly	AAG Lys	GGT Gly	TTG Leu 40	GAA Glu	TGG Trp	GTT Val	GCT Ala	CGC Arg 45	ATA Ile	AGA Arg	AGT Ser	144
AAA Lys	AGT Ser 50	AAT Asn	AAT Asn	TAT Tyr	GCA Ala	ACA Thr 55	TAT Tyr	TAT Tyr	GCC Ala	GAT Asp	TCA Ser 60	GTG Val	AAA Lys	GAC Asp	AGG Arg	192
TTC Phe 65	ACC Thr	ATC Ile	TCC Ser	AGA Arg	GAT Asp 70	GAT Asp	TCA Ser	CAA Gln	AGC Ser	ATG Met 75	CTC Leu	TAT	CTG Leu	CAA Gln	ATG Met 80	240 ·
AAC Asn	AAC Asn	TTG Leu	Lys	ACT Thr 85	GAG Glu	GAC Asp	ACA Thr	GCC Ala	ATG Met 90	TAT Tyr	TAC Tyr	TGT Cys	GTG Val	AGA Arg 95	CGG Arg	288
TTT Phe	GCT Ala	TAC Tyr	TGG Trp 100	GCC Ala	CAA Gln	GGG Gly	ACT Thr	CTG Leu 105	GTC Val	ACT Thr	GTC Val	TCT Ser	GCA Ala 110			330

(2) INFORMATION FOR SEQ ID NO:24:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 110 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
 (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

(2) INFORMATION FOR SEQ ID NO:25:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 357 base pairs
 (B) TYPE: nucleic acid

 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE: (ix) FEATURE:
- - (A) NAME/KEY: Coding Sequence (B) LOCATION: 1...357

 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

	GTT Val								48
	ATG Met								96
	ATG Met								 144
_	GAA Glu 50	 		 	 	 		 	 192
	AAA Lys	 		 	 				 240
	CTG Leu	 			 				 288
	ACC Thr	 	_					_	 336
	CTG Leu	 		 					357

- (2) INFORMATION FOR SEQ ID NO:26:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 119 amino acids

 - (B) TYPE: amino acid(C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

Glu Val Lys Leu Glu Glu Ser Gly Gly Gly Leu Val Gln Pro Jly Gly Ser Met Lys Leu Ser Cys Val Ala Ser Gly Phe Thr Phe Ser Asn Tyr 20 25 Trp Met Asn Trp Val Arg Gln Ser Pro Glu Lys Gly Leu Glu Trp Val 40 Ala Glu Val Arg Leu Lys Ser Asn Tyr Ala Thr His Tyr Ala Glu Ser 55 Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asp Ser Lys Ser Ser Val 75 70 Tyr Leu Gln Met Asn Asn Leu Arg Ala Glu Asp Thr Gly Ile Tyr Tyr 90 Cys Thr Arg Tyr Gly Arg Glu Gly Gly Val Ala Tyr Trp Gly Gln Gly 105 100 Thr Leu Val Thr Val Ser Ala 115

(2) INFORMATION FOR SEQ ID NO:27:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 360 base pairs

 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE: (ix) FEATURE:
- - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...360
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

_						GTT Val			48
						ACT Thr			96
			 	 		GGG Gly			144
		 	 	 		 ACA Thr 60	 	 	192
						GAT Asp			240
						GAC Asp			288
						GCT Ala			336
_	_		 GTC Val	_					360

(2) INFORMATION FOR SEQ ID NO:28:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 120 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

Glu Val Lys Thr Glu Glu Ser Gly Gly Gly Leu Val Gln Pro Gly Gly 1 15 Ser Met Lys Leu Ser Cys Val Ala Ser Gly Phe Thr Phe Ser Asn Tyr 20 25 Trp Met Asn Trp Val Arg Gln Ser Pro Glu Lys Gly Leu Glu Trp Val 35 40 Ala Glu Ile Arg Leu Lys Ser Asn Asn Tyr Ala Thr His Tyr Ala Glu Ser Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asp Ser Lys Ser Ser 70 75 Val Tyr Leu Gln Met Asn Asn Leu Arg Ala Glu Asp Thr Gly Ile Tyr 85 90 Tyr Cys Thr Arg Tyr Gly Arg Glu Gly Gly Phe Ala Tyr Trp Gly Glu 100 105 Gly Thr Leu Val Thr Val Ser Ala 115 120

(2) INFORMATION FOR SEQ ID NO:29:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 321 base pairs
 - (B) TYPE: nucleic acid (C) STRANDEDNESS: single

 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence (B) LOCATION: 1...321

 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

ACA Thr								GGA Gly	48
AAA Lys									96
AAC Asn								 	144
GAA Glu 50								 	192
GGC Gly								 	240
 GAT Asp	_	_		 	_			 	289
TTC Phe									321

(2) INFORMATION FOR SEQ ID NO:30:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 107 amino acids
 (B) TYPE: amino acid

 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

Glu Thr Thr Val Thr Gln Ser Pro Ala Ser Leu Ser Val Ala Thr Gly Glu Lys Val Thr Ile Arg Cys Ile Thr Ser Thr Asp Ile Asp Asp 20 25 Met Asn Trp Tyr Gln Gln Lys Pro Gly Glu Pro Pro Lys Leu Leu Ile 40 45 Ser Glu Gly Asn Thr Leu Arg Pro Gly Val Pro Ser Arg Phe Ser Ser 55 Ser Gly Tyr Gly Thr Asp Phe Val Phe Thr Ile Glu Asn Thr Leu Ser 70 75 Glu Asp Val Ala Asp Tyr Tyr Cys Leu Gln Ser Asp Asn Met Pro Leu 90 Thr Phe Gly Ala Gly Thr Lys Leu Glu Leu Lys

(2) INFORMATION FOR SEQ ID NO:31:

- (i) SEQUENCE CHARACTERISTICS:

 - (A) LENGTH: 321 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence (B) LOCATION: 1...321

 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

										ATG Met						48	}
GAC Asp	AGG Arg	GTC Val	AGC Ser 20	GTC Val	ACC Thr	TGC Cys	AAG Lys	GCC Ala 25	AGT Ser	CAG Gln	AAT Asn	GTG Val	GGT Gly 30	ACT Thr	AAT Asn	96	į
										TCT Ser						144	
TAC	TCG Ser 50	GCA Ala	TCC Ser	TAC Tyr	CGG Arg	TAC Tyr 55	AGT Ser	GGA Gly	GTC Val	CCT Pro	GAT Asp 60	CGC Arg	TTC Phe	ACA Thr	GGC	192	
										ATC Ile 75						240	i
										TAT Tyr						288	ŕ
					ACC Thr					AAA Lys						32	1

(2) INFORMATION FOR SEQ ID NO:32:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 107 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

Asp Ile Val Met Thr Gln Ser Gln Lys Phe Met Ser Thr Ser Val Gly 10 Asp Arg Val Ser Val Thr Cys Lys Ala Ser Gln Asn Val Gly Thr Asn 20 Val Ala Trp Tyr Gln Gln Lys Pro Gly Gln Ser Pro Lys Ala Leu Ile 35 45 40 Tyr Ser Ala Ser Tyr Arg Tyr Ser Gly Val Pro Asp Arg Phe Thr Gly 55 60 Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Asp Val Gln Ser 70 75 Glu Asp Leu Ala Glu Tyr Phe Cys Gln Gln Tyr Asn Ile Tyr Pro Leu 85 90 Thr Phe Gly Ala Gly Thr Lys Leu Glu Leu Lys 100

(2) INFORMATION FOR SEQ ID NO:33:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 321 base pairs

 - (B) TYPE: nucleic acid (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (iii) HYPCTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:
- (ix) FEATURE:
 - (A) NAMI/KEY: Coding Sequence (B) LOCATION: 1...321

 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

		 	 	 			GGA Gly		48
						GAT Asp 30			96
						CTC Leu			144
						TTC Phe			192
						ACG Thr			240
						ATG Met		;	288
 TTC Phe	 	 	 	 _					321

(2) INFORMATION FOR SEQ ID NO:34:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 107 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

Glu Thr Thr Val Thr Gln Ser Pro Ala Ser Leu Ser Val Ala Thr Gly 10 Glu Lys Val Thr Ile Arg Cys Ile Thr Ser Thr Asp Ile Asp Asp 25 20 Met Asn Trp Tyr Gln Gln Lys Pro Gly Glu Pro Pro Lys Leu Leu Ile 35 45 Ser Glu Gly Asn Thr Leu Arg Pro Gly Val Pro Ser Arg Phe Ser Ser 55 Ser Gly Tyr Gly Thr Asp Phe Val Phe Thr Ile Glu Asn Thr Leu Ser 75 Glu Asp Val Ala Asp Tyr Tyr Cys Leu Gln Ser Asp Asn Met Pro Phe 85 90 Thr Phe Gly Ser Gly Thr Lys Leu Glu Ile Lys

(2) INFORMATION FOR SEQ ID NO:35:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 345 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single

 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence (B) LOCATION: 1...345

 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

_			CTT Leu							48
			CTC Leu 20			 			 	96
			TGG Trp							144
		_	AGA Arg	 	 	 			 	192
_	_		GAC Asp					_		240
			CAA Gln	 	 	 		 	 	288
			AGA Arg 100							336
	TCC Ser									345

(2) INFORMATION FOR SEQ ID NO:36:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 115 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

Glu Val Gln Leu Val Glu Ser Gly Gly Gly Leu Val Gln Pro Lys Gly 10 Ser Leu Lys Leu Ser Cys Ala Ala Ser Gly Phe Thr Phe Asn Thr Tyr 20 25 Ala Met Asn Trp Val Arg Gln Leu Gln Gly Lys Gly Leu Glu Trp Val 35 40 45 Ala Arg Ile Arg Ser Lys Ser Asn Asn Tyr Ala Thr Tyr Tyr Ala Asp 55 60 Ser Val Lys Asp Arg Phe Thr Ile Ser Arg Asp Asp Ser Gln Ser Met 70 75 Leu Tyr Leu Gln Met Asn Asn Leu Lys Thr Glu Asp Thr Ala Met Tyr 85 90 Tyr Cys Val Arg Arg Arg Asp Tyr Trp Gly Gln Gly Thr Ser Val Thr Val Ser Ser 115

(2) INFORMATION FOR SEQ ID NO:37:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 372 base pairs (B) TYPE: nucleic acid

 - (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence (B) LOCATION: 1...372 (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

		 	GAG Glu	 				 	48
		 	TGT Cys	 	 		 	 	96
		 	CGC Arg	 	 				144
			AAA Lys						192
			TTC Phe 70						240
	 	 	AAC Asn	 	 				288
			ATC Ile				_		336
			ACC Thr						372

(2) INFORMATION FOR SEQ ID NO:38:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 124 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

Glu Val Lys Leu Val Glu Ser Gly Gly Gly Leu Val Gln Pro Gly Gly 1 10 Ser Leu Arg Leu Ser Cys Ala Thr Ser Gly Phe Thr Phe Thr Asp Tyr 25 Tyr Met Ser Trp Val Arg Gln Pro Pro Gly Lys Ala Leu Glu Trp Leu 40 Gly Leu Ile Arg Asn Lys Ala Asn Gly Tyr Thr Thr Glu Tyr Ser Ala 55 60 Ser Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn Ser Gln Ser Ile 75 70 Leu Tyr Leu Gln Met Asn Thr Leu Arg Ala Glu Asp Ser Ala Thr Tyr 90 Tyr Cys Ala Arg Asp Ile Tyr Tyr Asp Tyr Asp Tyr Tyr Ala Met Asp 100 105 Tyr Trp Gly Gln Gly Thr Ser Val Thr Val Ser Ser

(2) INFORMATION FOR SEQ ID NO:39:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 111 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: Genomic DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:
- (ix) FEATURE:
 - (A) NAME/KEY: Coding Sequence
 - (B) LOCATION: 1...111
 - (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

CAC TGG AAC CAA CAG AAA CCA GGA CAG CCA CCC AGA CTC CTC ATC TAT

His Trp Asn Gln Gln Lys Pro Gly Gln Pro Pro Arg Leu Leu Ile Tyr

1 5 10 15

CTT GTA TCC AAC CTA GAA TCT GGG GTC CCT GCC AGG TTC AGT GGC AGT 96
Leu Val Ser Asn Leu Glu Ser Gly Val Pro Ala Arg Phe Ser Gly Ser
20 25 30

GGG TCT GGG ACA GAC Gly Ser Gly Thr Asp 35 111

- (2) INFORMATION FOR SEQ ID NO:40:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 37 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

His Trp Asn Gln Gln Lys Pro Gly Gln Pro Pro Arg Leu Leu Ile Tyr 10 Leu Val Ser Asn Leu Glu Ser Gly Val Pro Ala Arg Phe Ser Gly Ser 20 25 Gly Ser Gly Thr Asp 35

- (2) INFORMATION FOR SEQ ID NO:41:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 30 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:

GAAGATCTAG ACTTACTATG CAGCATCAGC

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- (2) INFORMATION FOR SEQ ID NO:42:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 34 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

AGGAGACTAG TGGTTACTAA TTTGGGAAGG ACTG

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- (2) INFORMATION FOR SEQ ID NO:43: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 48 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTISENSE: NO (v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE: (xi) SEQUENCE DESCRIPTION: SEQ ID NO:43: GAGACGACTA GTGGTGGCGG TGGCTCTCCA TTCGTTTGTG AATATCAA 48 (2) INFORMATION FOR SEQ ID NO:44: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 40 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTISENSE: NO (v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE: (xi) SEQUENCE DESCRIPTION: SEQ ID NO:44: TTACTAGCTA GCATAATAAC GGAATACCCA AAAGAACTGG 40 (2) INFORMATION FOR SEQ ID NO:45: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 36 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO

 - (iv) ANTISENSE: NO
 - (v) FRAGMENT TYPE:
 - (vi) ORIGINAL SOURCE:
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:

TATGCTAGCT AGTAACACGA GAGGTTTCCC GACTGG

- (2) INFORMATION FOR SEQ ID NO:46:
- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 30 base pairs

 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE:
- (vi) ORIGINAL SOURCE:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:46:

AGCTTTGAAT TCGTGAAATT GTTATCCGCT

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INDICATIONS RELATING TO A DEPOSITED MICROORGANISM

(PCT Rule 13bis)

A. The indications made below relate to the microorganism related to the	ferred to in the description
on page 17 , line	
B. IDENTIFICATION OF DEPOSIT	Further deposits are identified on an additional sheet
Name of depositary institution	
American Type Culture Collectio	on .
Address of depositary institution (including postal code and co	untry)
12301 Parklawn Drive Rockville, Maryland 20852 United States of America	
Date of deposit	Accession Number
14 April 1993 (16.04.93)	HB 11330
C. ADDITIONAL INDICATIONS (leave blank if not applications)	able) This information is continued on an additional sheet
	ARE MADE (if the indications are not for all designated States)
E. SEPARATE FURNISHING OF INDICATIONS (leave b	
The indications listed below will be submitted to the Internations Number of Deposit")	al Bureau later (specify the general nature of the indications e.g., "Accession
For receiving Office use only	For International Bureau use nly
This sheet was received with the international application	This sheet was received by the International Bureau on:
Auth rized officer	Authorized fficer

WHAT IS CLAIMED IS:

- 1. An isolated nucleic acid sequence coding for a heavy chain Fd fragment of a monoclonal antibody,

 5 wherein the monoclonal antibody immunoreacts with a lead cation, and wherein the nucleic acid sequence includes the sequence for the heavy chain variable region from monoclonal antibody 6B11 (SEQ ID NO:1),

 1254 (SEQ ID NO:3), 8E7 (SEQ ID NO:5), 10G5 (SEQ ID NO:7), 14F11 (SEQ ID NO:9), 13D10 (SEQ ID NO:17),

 11D11 (SEQ ID NO:19), 14G11 (SEQ ID NO:21), 6F5 (SEQ ID NO:23), 7D10 (SEQ ID NO:25), 4E8 (SEQ ID NO:27), 2B4 (SEQ ID NO:35), or 2E7 (SEQ ID NO:37).
- 15 2. An isolated nucleic acid sequence coding for a light chain of a monoclonal antibody, wherein the monoclonal antibody immunoreacts with a lead cation, and wherein the nucleic acid sequence includes the sequence for the light chain variable region from monoclonal antibody 6B11 (SEQ ID NO:11), 1254 (SEQ ID NO:13), 14F11 (SEQ ID NO:15), 10G5 (SEQ ID NO:29), 2E7 (SEQ ID NO:31), or 7D10 (SEQ ID NO:33).
- 25 3. An expression cassette comprising:

a nucleic acid sequence coding for a heavy chain Fd fragment of a monoclonal antibody, wherein the monoclonal antibody immunoreacts with a lead cation, wherein the nucleic acid sequence includes the sequence for the heavy chain variable region from monoclonal antibody 6B11 (SEQ ID NO:1), 1254 (SEQ ID NO:3), 8E7 (SEQ ID NO:5), 10G5 (SEQ ID NO:7), 14F11 (SEQ ID NO:9), 13D10 (SEQ ID NO:17), 11D11 (SEQ ID NO:19), 14G11 (SEQ ID NO:21), 6F5 (SEQ ID NO:23), 7D10 (SEQ ID NO:25), 4E8 (SEQ ID NO:27), 2B4 (SEQ ID NO:35), or 2E7 (SEQ ID NO:37);

wherein the nucleic acid sequence coding for the heavy chain Fd fragment is operably linked to a promoter functional in a vector.

An expression cassette according to claim 3 further comprising a leader sequence located between the promoter and the nucleic acid sequence, wherein the leader sequence functions to direct the heavy chain Fd fragment to a membrane in a host cell.

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- 5. An expression cassette comprising:
 - (a) a nucleic acid sequence coding for a light chain for a monoclonal antibody, wherein the monoclonal antibody immunoreacts with a lead cation, wherein the nucleic acid sequence includes the sequence for the light chain variable region from monoclonal antibody 6B11 (SEQ ID NO:11), 1254 (SEQ ID NO:13), 14F11 (SEQ ID NO:15), 10G5 (SEQ ID

NO:29), 2E7 (SEQ ID NO:31), or 7D10 (\$EQ ID NO:33),

- wherein the nucleic acid sequence coding for the light chain is operably linked to a promoter functional in a vector.
- An expression cassette according to claim 5 further comprising a leader sequence located between the promoter and the nucleic acid sequence, wherein the leader sequence functions to direct the light chain to a membrane in a host cell.
- 30 7. An expression cassette coding for a fusion protein comprising:
 - (a) a first nucleic acid sequence coding for a heavy chain Fd fragment of a monoclonal antibody, wherein the monoclonal antibody reacts with lead cation, wherein the first nucleic acid sequence includes the sequence for the heavy chain variable region from monoclonal antibody 6B11 (SEQ

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ID NO:1), 1254 (SEQ ID NO:3), 8E7 (SEQ ID NO:5), 10G5 (SEQ ID NO:7), 14F11 (SEQ ID NO:9), 13D10 (SEQ ID NO:17), 11D11 (SEQ ID NO:19), 14G11 (SEQ ID NO:21), 6F5 (SEQ ID NO:23), 7D10 (SEQ ID NO:25), 4E8 (SEQ ID NO:27), 2B4 (SEQ ID NO:35), or 2E7 (SEQ ID NO:37), and wherein the first nucleic acid sequence is linked for co-expression to

- (b) a second nucleic acid sequence coding for a phage coat protein or a portion thereof, thereby forming a nucleic acid sequence encoding the fusion protein, wherein the fusion protein includes the heavy chain Fd fragment fused to the phage coat protein or portion thereof; and
- (c) a promoter that is functional in a vector, wherein the promoter is operably linked to the first and second nucleic acid sequences and provides for expression of the fusion protein.
- 8. An expression cassette according to claim 7 further comprising a leader sequence located between the promoter and the nucleic acid sequence coding for the fusion protein, wherein the leader sequence directs expression of the fusion protein to a membrane of a host cell.

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9. An expression cassette comprising:

(a) a first nucleic acid sequence coding for a heavy chain Fd fragment of a monoclonal antibody, wherein the monoclonal antibody immunoreacts with a lead cation, wherein the first nucleic acid sequence includes the sequence for the heavy chain variable region from monoclonal antibody 6B11 (SEQ ID NO:1), 1254 (SEQ ID NO:3), 8E7 (SEQ ID NO:5), 10G5 (SEQ ID NO:7), 14F11 (SEQ ID NO:9), 13D10 (SEQ ID NO:17), 11D11 (SEQ ID NO:19), 14G11 (SEQ ID NO:21), 6F5 (SEQ ID NO:23),

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7D10 (SEQ ID NO:25), 4E8 (SEQ ID NO:27), 2B4 (SEQ ID NO:35), or 2E7 (SEQ ID NO:37), and

wherein the first nucleic acid sequence is linked for co-expression to

- (b) a second nucleic acid sequence for phage coat protein or a portion thereof to form a nucleic acid sequence encoding a fusion protein, wherein the fusion protein includes the heavy chain Fd fragment fused to the phage coat protein or portion thereof;
- (c) a first promoter that is functional in a vector, wherein the first promoter is operably linked to the nucleic acid sequence encoding the fusion protein and provides for expression of the fusion protein; and
- (d) a fourth nucleic acid sequence coding for a light chain of an antibody;

 wherein the fourth nucleic acid sequence is operably linked to a second promoter functional in the vector, and wherein the first and second promoters are coordinately expressed.
- 10. The expression cassette of claim 9, wherein the light chain is a light chain of a monoclonal antibody which immunoreacts with a lead cation, and wherein the fourth nucleic acid sequence includes the sequence for the light chain variable region from monoclonal antibody 6B11 (SEQ ID NO:11), 1254 (SEQ ID NO:13), 14F11 (SEQ ID NO:15), 10G5 (SEQ ID NO:29), 2E7 (SEQ ID NO:31), or 7D10 (SEQ ID NO:33).
 - 11. A phagemid vector having the expression cassette of claim 9.
 - 12. A metal binding polypeptide comprising an amino acid sequence for a heavy chain Fd fragment from a

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monoclonal antibody, wherein the monoclonal antibody immunoreacts with a lead cation, and wherein the amino acid sequence for the heavy chain Fd fragment includes the sequence for the heavy chain variable region from monoclonal antibody 6B11 (SEQ ID NO:2), 1254 (SEQ ID NO:4), 8E7 (SEQ ID NO:6), 10G5 (SEQ ID NO:8), 14F11 (SEQ ID NO:10), 13D10 (SEQ ID NO:18), 11D11 (SEQ ID NO:20), 14G11 (SEQ ID NO:22), 6F5 (SEQ ID NO:24), 7D10 (SEQ ID NO:26), 4E8 (SEQ ID NO:28), 2B4 (SEQ ID NO:36), or 2E7 (SEQ ID NO:38).

- 13. The polypeptide of claim 12 further comprising a heavy chain Fc fragment fused to the heavy chain Fd fragment.
 - 14. The polypeptide of claim 12 further comprising a phage coat protein or portion thereof fused to the heavy chain Fd fragment.

15. A heavy chain of a monoclonal antibody, wherein the monoclonal antibody immunoreacts with a lead cation, and wherein the heavy chain has an amino acid sequence which includes the heavy chain variable region from monoclonal antibody 6B11 (SEQ ID NO:2), 1254 (SEQ ID NO:4), 8E7 (SEQ ID NO:6), 10G5 (SEQ ID NO:8), 14F11 (SEQ ID NO:10), 13D10 (SEQ ID NO:18), 11D11 (SEQ ID NO:20), 14G11 (SEQ ID NO:22), 6F5 (SEQ ID NO:24), 7D10 (SEQ ID NO:26), 4E8 (SEQ ID NO:28), 2B4 (SEQ ID NO:36), or 2E7 (SEQ ID NO: 38).

16. A fusion protein comprising a heavy chain Fd fragment of a monoclonal antibody and a phage coat protein or portion thereof, wherein the monoclonal antibody immunoreacts with a lead cation.

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- 17. A phage comprising a fusion protein according to claim 16 and a light chain of an antibody.
- 18. The phage of claim 17 wherein the light chain is a light chain of a monoclonal antibody which immunoreacts with the lead cation.
- 19. A metal binding polypeptide comprising an amino acid sequence for a light chain from a monoclonal antibody, wherein the monoclonal antibody immunoreacts with a lead cation, and wherein the light chain amino acid sequence includes the sequence for the light chain variable region from monoclonal antibody 6B11 (SEQ ID NO:12), 1254 (SEQ ID NO:14), 14F11 (SEQ ID NO:16), 10G5 (SEQ ID NO:34).
- 20. A light chain of a monoclonal antibody, wherein the monoclonal antibody immunoreacts with a lead cation, and wherein the light chain has an amino acid sequence which includes the sequence for the light chain variable region from monoclonal antibody 6B11 (SEQ ID NO:12), 1254 (SEQ ID NO:14), 14F11 (SEQ ID NO:16), 10G5 (SEQ ID NO:30), 2E7 (SEQ ID NO:32), or 7D10 (SEQ ID NO:34).
- 21. A recombinantly produced Fab fragment that immunoreacts with a lead cation, wherein the Fab fragment has a heavy chain Fd fragment with an amino acid sequence which includes the sequence for the heavy chain variable region from monoclonal antibody 6B11 (SEQ ID NO:2), 1254 (SEQ ID NO:4), 8E7 (SEQ ID NO:6), 10G5 (SEQ ID NO:8), 14F11 (SEQ ID NO:10), 13D10 (SEQ ID NO:18), 11D11 (SEQ ID NO:20), 14G11 (SEQ ID NO:22), 6F5 (SEQ ID NO:24), 7D10 (SEQ ID NO:26), 4E8 (SEQ ID NO:28), 2B4 (SEQ ID NO:36), or 2E7 (SEQ ID NO: 38).

- 22. A recombinantly produced Fab fragment that immunoreacts with a lead cation, the Fab fragment comprising a light chain having an amino acid sequence which includes the sequence for the light chain variable region from monoclonal antibody 6B11 (SEQ ID NO:12), 1254 (SEQ ID NO:14), 14F11 (SEQ ID NO:16), 10G5 (SEQ ID NO:30), 2E7 (SEQ ID NO:32), or 7D10 (SEQ ID NO:34).
- 10 23. A purified antibody comprising a Fab fragment that immunoreacts with a lead cation, the Fab fragment comprising a heavy chain Fd fragment having an amino acid sequence which includes the sequence for the heavy chain variable region from monoclonal antibody 6B11 (SEQ ID NO:2), 1254 (SEQ ID NO:4), 8E7 (SEQ ID NO:6), 10G5 (SEQ ID NO:8), 14F11 (SEQ ID NO:10), 13D10 (SEQ ID NO:18), 11D11 (SEQ ID NO:20), 14G11 (SEQ ID NO:22), 6F5 (SEQ ID NO:24), 7D10 (SEQ ID NO:26), 4E8 (SEQ ID NO:28), 2B4 (SEQ ID NO:36), or 2E7 (SEQ ID NO:38).
 - 24. The purified antibody of claim 23 which is a recombinantly produced monoclonal antibody.
- 25 25. A purified antibody comprising a Fab fragment that immunoreacts with a lead cation, the Fab fragment comprising a light chain having an amino acid sequence which includes the sequence for the light chain variable region from monoclonal antibody 6B11 (SEQ ID NO:12), 1254 (SEQ ID NO:14), 14F11 (SEQ ID NO:16), 10G5 (SEQ ID NO:30), 2E7 (SEQ ID NO:32), or 7D10 (SEQ ID NO:34).
- 26. A kit for detecting the presence of lead cations
 comprising a metal binding polypeptide which
 includes an amino acid sequence for a heavy chain
 Fd fragment from a monoclonal antibody, wherein the

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monoclonal antibody immunoreacts with a lead cation, and the heavy chain Fd fragment has an amino acid sequence which includes the sequence for the heavy chain variable region from monoclonal antibody 6B11 (SEQ ID NO:2), 1254 (SEQ ID NO:4), 8E7 (SEQ ID NO:6), 10G5 (SEQ ID NO:8), 14F11 (SEQ ID NO:10), 13D10 (SEQ ID NO:18), 11D11 (SEQ ID NO:20), 14G11 (SEQ ID NO:22), 6F5 (SEQ ID NO:24), 7D10 (SEQ ID NO:26), 4E8 (SEQ ID NO:28), 2B4 (SEQ ID NO:36), or 2E7 (SEQ ID NO:38).

- 27. A kit for detecting the presence of lead cations comprising a metal binding polypeptide which includes an amino acid sequence for a light chain from a monoclonal antibody, wherein the monoclonal antibody immunoreacts with a lead cation, and the light chain has an amino acid sequence including the sequence for the light chain variable region from monoclonal antibody 6B11 (SEQ ID.NO:12), 1254 (SEQ ID NO:14), 14F11 (SEQ ID NO:16), 10G5 (SEQ ID NO:30), 2E7 (SEQ ID NO:32), or 7D10 (SEQ ID NO:34).
 - 28. A method of producing a metal binding polypeptide comprising:
- i) expressing a nucleic acid sequence in a transformed host cell to form a lead binding polypeptide, said nucleic acid sequence comprises a sequence coding for a heavy chain Fd fragment of a monoclonal antibody which is capable of immunoreacting with a lead cation, wherein the sequence coding for the heavy chain Fd fragment includes a sequence for a heavy chain variable region from monoclonal antibody 6B11 (SEQ ID NO:1), 1254 (SEQ ID NO:3), 8E7 (SEQ ID NO:5), 10G5 (SEQ ID NO:7), 14F11 (SEQ ID NO:9), 13D10 (SEQ ID NO:18), 11D11 (SEO ID NO:20), 14G11 (SEQ ID NO:22), 6F5

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(SEQ ID NO:24), 7D10 (SEQ ID NO:26), 4E8 (SEQ ID NO:28), 2B4 (SEQ ID NO:36), or 2E7 (SEQ ID NO:38).

- 29. The method of claim 28 wherein the metal binding polypeptide is a fusion protein and the sequence coding for the heavy chain Fd fragment is linked for co-expression to a nucleic acid sequence coding for an adjunct peptide.
- 10 30. A method of producing a metal binding polypeptide comprising:

expressing a nucleic acid sequence in a transformed host cell to form a lead binding polypeptide, said nucleic acid sequence comprising a sequence coding for a light chain of a monoclonal antibody which is capable of immunoreacting with a lead cation, wherein the sequence encoding for the light chain includes a sequence encoding a light chain variable region from monoclonal antibody 6B11 (SEQ ID NO:11), 1254 (SEQ ID NO:13), 14F11 (SEQ ID NO:15), 10G5 (SEQ ID NO:29), 2E7 (SEQ ID NO:31), or 7D10 (SEQ ID NO:33).

31. A metal binding polypeptide obtainable by a process comprising:

expressing a nucleic acid sequence in a transformed host cell, wherein the nucleic acid sequence consists essentially of a sequence coding for a heavy chain variable region from monoclonal antibody 6B11 (SEQ ID NO:1), 1254 (SEQ ID NO:3), 8E7 (SEQ ID NO:5), 10G5 (SEQ ID NO:7), 14F11 (SEQ ID NO:9), 13D10 (SEQ ID NO:17), 11D11 (SEQ ID NO:19), 14G11 (SEQ ID NO:21), 6F5 (SEQ ID NO:23), 7D10 (SEQ ID NO:25), 4E8 (SEQ ID NO:27), 2B4 (SEQ ID NO:35), or 2E7 (SEQ ID NO:37).

32. A method of producing a metal binding antibody specific for a metallic cation comprising injecting

an antibody capable of immunoreacting with the metallic cation into a mammal.

- 33. The method of claim 32 wherein the metallic cation is a cation of lead or mercury.
- 34. A hybridoma of myeloma immortal cells and mammalian immune cells from a mammal previously immunized with a first antibody capable of immunoreacting with a metallic cation, wherein the hybridoma produces a second monoclonal antibody capable of immunoreacting with the metallic cation in an exposed state.
- 15 35. A lead binding polypeptide comprising a light chain of a monoclonal antibody capable of immunoreacting with a lead cation.
- 36. The polypeptide of claim 35 wherein the light chain is a member of the V_k 32 light chain family.
- 37. The polypeptide of claim 35 wherein the light chain comprises a motifof five consecutive amino acid residues, where four of the five amino acid residues each have a carboxylic acid-containing side chain.
 - 38. The polypeptide of claim 37 wherein the motif is present in a CDR1 region of the light chain.
 - 39. The polypeptide of claim 38 wherein the motif is positioned at amino acid residues 28, 29, 30 31 and 32.
- 35 40. The polypeptide of claim 37 wherein the amino acid residues having a carboxylic acid-containing side chain are glutamic acid or aspartic acid.

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41. An isolated nucleic acid sequence coding for a light chain of a monoclonal antibody capable of immunoreacting with a lead cation, wherein the monoclonal antibody is a member of the V_k 32 light chain family.

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Sequences of Heavy Chains of Lead-Specific Antibodies

Sequences of Light Chain Variable Regions of Lead-Specific Antibodies

٦ ا Lead-Specific Heavy Chain Sequences

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FIG. 5

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s TCA	C TGT	a gca	a GCC	s TCT	g gga	F TTC	T ACC	P TTC	NI	T	Y	A	M	N	W	v	R CGC	Q CAG	40 L CTC
Q CAA	G GGA	K AAG	G GGT	L TTG	e Gaa	w Tgg	V GTT	a GCT	D	T	52 R AGA	S	K	S	N	n aat	Y TAT	A GCA	T ACA
Y TAT	Y	A GCC		S TCA	V GTG	K AAA	D GAC	R AGG	P TTC	T	I	70 S TCC	R	D	D GAT	S TCA	Q CAA	s AGC	m atg
L	Y TAT	BO L	Q CAJ	M ATG	N RAC	N RAC	L TTG	K : AAA	T ACI	E E	D GAC	T ACA	A GCC	M ATG	90 Y TAT	Y TAC	C TGT	v GTG	R AGA
R	96 R	<u> </u>					Y	_	G G GGT	0	G A GGA	T	S	v	T	V	S TCC	S TCA	

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FIG. 6

2E7 heavy chain

Glu Val Lys Leu Val Glu Ser Gly Gly Gly Leu Val Gln Pro Gly Gly Ser Leu Arg Leu GAG GTG AAG CTG GTG GAG TCT GGA GGA GGC TTG GTA CAG CCT GGG GGT TCT CTG AGA CTC

Ser Cys Ala Thr Ser Gly Phe Thr Phe Thr Asp Tyr Tyr Met Ser Trp Val Arg Gln Pro
TCC TGT GCA ACT TCT GGG TTC ACC TTC ACT GAT TAC TAC ATG AGC TGG GTC CGC CAG CCT

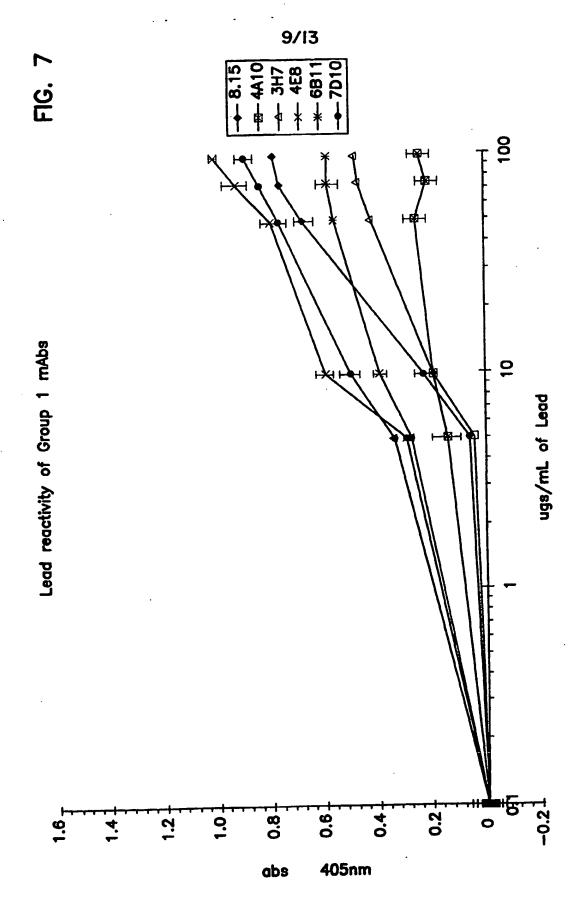
Pro Gly Lys Ala Leu Glu Trp Leu Gly Leu Ile Arg Asn Lys Ala Asn Gly Tyr Thr Thr
CCA GGA AAG GCA CTT GAG TGG TTG GGT TTG ATT AGA AAC AAA GCT AAT GGT TAC ACA ACA

Glu Tyr Ser Ala Ser Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn Ser Gln Ser Ile
GAG TAC AGT GCA TCT GTG AAG GGT CGG TTC ACC ATC TCC AGA GAT AAT TCC CAA AGC ATC

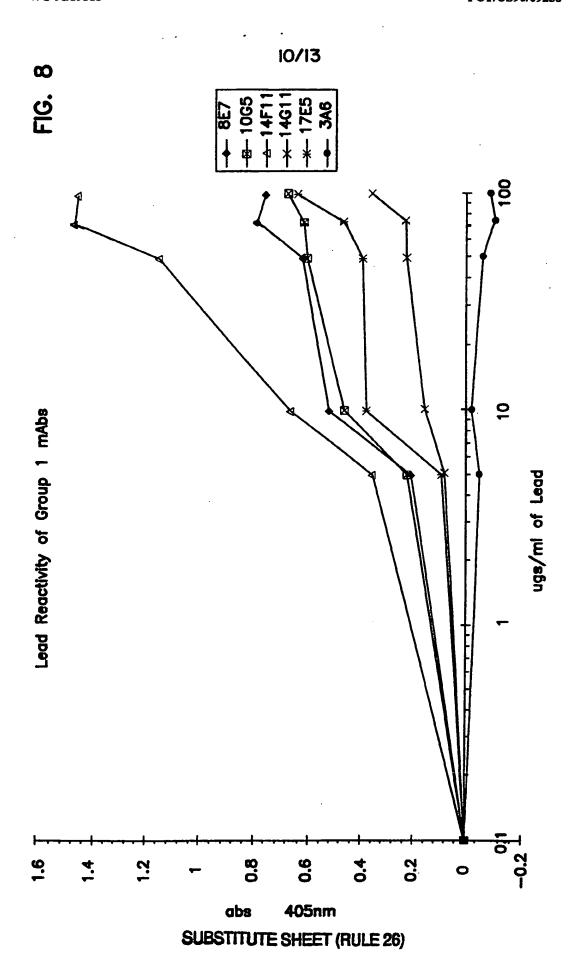
Leu Tyr Leu Gln Met Asn Thr Leu Arg Ala Glu Asp Ser Ala Thr Tyr Tyr Cys Ala Arg
CTC TAT CTT CAA ATG AAC ACC CTG AGA GCT GAG GAC AGT GCC ACT TAT TAC TGT GCA AGA

Asp Ile Tyr Tyr Asp Tyr Asp Tyr Tyr Ala Met Asp Tyr Trp Gly Gln Gly Thr Ser Val
GAT ATC TAC TAT GAT TAC GAC TAC TAT GCT ATG GAC TAC TGG GGT CAA GGA ACC TCA GTC

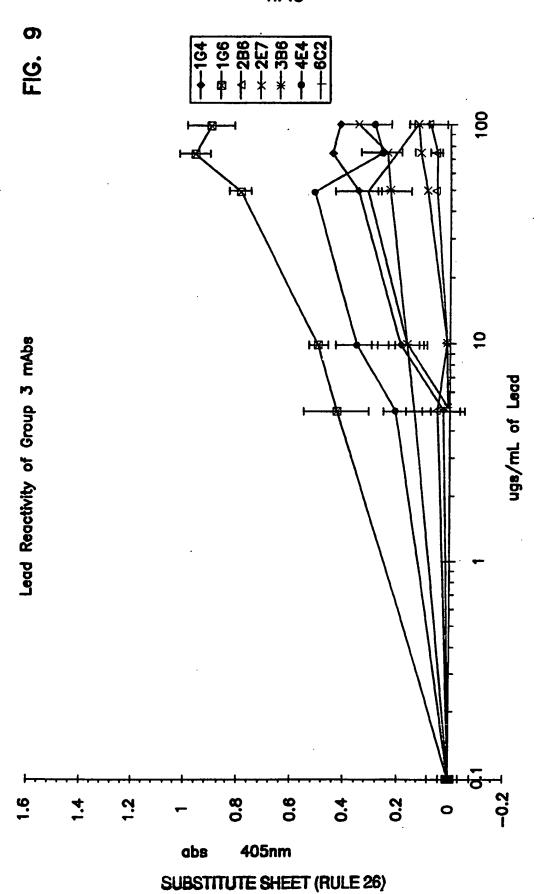
Thr Val Ser Ser
ACC GTC TCC TCA

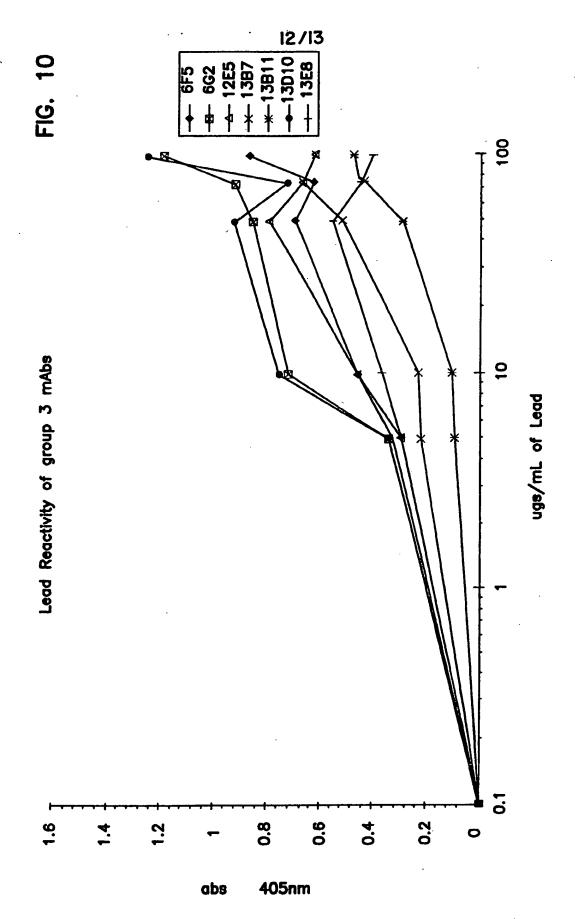


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